

**SOLID SUPPORT ASSAY SYSTEMS AND METHODS UTILIZING  
NON-STANDARD BASES**

**BACKGROUND OF THE INVENTION**

[0001] A variety of different methods have been developed to assay oligonucleotides, including DNA or RNA fragments. Such assays are typically directed to determining whether a sample includes oligonucleotides having a particular target oligonucleotide sequence. In some instances, oligonucleotide sequences differ by only a few nucleotides, as in the case of many allelic sequences. Single nucleotide polymorphisms (SNPs) refer to alleles that differ by a single nucleotide. Even this single nucleotide difference can, at least in some instances, change the associated genetic response or traits. Accordingly, to determine which allele is present in a sample, the assay technique must be sufficiently sensitive to distinguish between closely related sequences.

[0002] Many assay techniques include multiple components, each of which hybridizes to other component(s) in the assay. Non-specific hybridization between components (i.e., the hybridization of two non-complementary sequences) produces background noise in the assay. For example, closely related, but not identical, sequences can form imperfect duplexes in which base pairing is interrupted at positions where the two single strands are not complementary. Non-specific hybridization increases when the hybridizing components have similar sequences, as would be the case, for example, for many alleles and particularly for SNP alleles. Thus, for example, hybridization assays to determine which allele is present in a sample would benefit from methods that reduce non-specific hybridization or reduce the impact of non-specific hybridization on the assay.

**BRIEF SUMMARY OF THE INVENTION**

[0003] Generally, the present invention relates to methods, kits, and compositions for assaying oligonucleotides. In addition, the invention relates to methods, kits, and compositions for assaying oligonucleotides using non-standard bases. One embodiment provides a method of assaying an analyte-specific sequence. A capture oligonucleotide comprising a molecular recognition sequence having at least one non-standard base coupled to a support (e.g., a single solid support, such as a chip or wafer, or a particulate support) is contacted with a sample under suitable hybridizing conditions to hybridize to

a target oligonucleotide, if present in the sample. The target oligonucleotide comprises a tagging sequence complementary to the molecular recognition sequence of the capture oligonucleotide and the analyte-specific sequence or a complement of the analyte-specific sequence. Hybridization of the target oligonucleotide to the capture oligonucleotide is detected.

[0004] Another embodiment provides another method of assaying an analyte-specific sequence. A capture oligonucleotide coupled to a support and comprising a molecular recognition sequence that is the same as or complementary to at least a portion of the analyte-specific sequence is contacted with a sample under hybridizing conditions to hybridize to a target oligonucleotide. The target oligonucleotide comprises a tagging sequence comprising at least one non-standard base and the analyte-specific sequence or a complement of the analyte-specific sequence. The capture oligonucleotide is enzymatically extended using the target oligonucleotide as a template and a complementary non-standard base is incorporated opposite the non-standard base of the tagging sequence. A reporter group is also incorporated into an extended portion of the capture oligonucleotide. Hybridization of the target oligonucleotide to the capture oligonucleotide is detected.

[0005] Yet another embodiment provides another method of assaying an analyte-specific sequence. An analyte having the analyte-specific sequence is contacted with a first primer and a second primer. The first primer comprises a tagging sequence and a sequence complementary to a first sequence of the analyte. The second primer comprises a sequence complementary to a second sequence of the analyte and a non-standard base. The first and second primers are enzymatically extended to form a target oligonucleotide and a second oligonucleotide, respectively. One of the target oligonucleotide and the second oligonucleotide comprises the analyte-specific sequence, and the other comprises a sequence complementary to the analyte-specific sequence. Extension of the first primer is substantially halted when the non-standard base of the second primer is encountered. A non-standard base complementary to the non-standard base of the second primer is incorporated into the extended first primer opposite the non-standard base of the second primer. A capture oligonucleotide molecular recognition sequence that is the same as or complementary to at least a portion of the analyte-specific sequence coupled to a support is contacted with the target oligonucleotide under hybridizing

conditions to hybridize to the target oligonucleotide comprising a tagging sequence and the analyte-specific sequence or complement thereof. Hybridization of the target oligonucleotide to the capture oligonucleotide is detected.

[0006] Other embodiments include kits for applying the methods described above. The kits include support(s) and capture oligonucleotides. The kits also include the target oligonucleotides or components for making the target oligonucleotides from an analyte. Such components can include, for example, a polymerase and first and second primers that are complementary to sequences of the analyte, where either the first or second primers include the tagging sequence. For some methods, the kit can also include a non-standard base or nucleotide triphosphate of a non-standard base for incorporation.

[0007] The above summary of the present invention is not intended to describe each disclosed embodiment or every implementation of the present invention. The Figures and the detailed description which follow more particularly exemplify these embodiments.

### **BRIEF DESCRIPTION OF THE DRAWINGS**

[0008] The invention may be more completely understood in consideration of the following detailed description of various embodiments of the invention in connection with the accompanying drawings, in which:

[0009] Figure 1 displays chemical structures for a number of non-standard bases, where A is the point of attachment to a polymeric backbone, X is N or C-Z, Y is N or C-H, and Z is H or a substituted or unsubstituted alkyl group;

[0010] Figures 2A and 2B schematically illustrate two examples of oligonucleotide hybridization to a solid support, according to the invention;

[0011] Figure 3 illustrates steps in a first assay for an analyte-specific sequence, according to the invention;

[0012] Figure 4 illustrates steps in a second assay for an analyte-specific sequence, according to the invention;

[0024] Figure 16 illustrates steps in a tenth assay for an analyte-specific sequence, according to the invention.

[0025] Figure 17 is a graph of results from an assay of alleles, according to the invention.

[0026] Although the invention is amenable to various modifications and alternative forms, specifics thereof have been shown by way of example in the drawings and will be described in detail. It should be understood, however, that the invention is not limited to the particular embodiments described. On the contrary, the intention is to cover all modifications, equivalents, and alternatives falling within the spirit and scope of the invention.

#### **DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT**

[0027] This application is related to U.S. Provisional Application Serial No. 60/240,397, filed October 14, 2000, U.S. Provisional Application Serial No 60/282,831, filed April 10, 2001, and U.S. Patent Application Serial No. 09/861,292, filed May 18, 2001, and U.S. Provisional Application Serial No. 60/293,259, filed May 22, 2001, all of which are incorporated herein by reference.

[0028] The present invention relates to assays and methods of assaying oligonucleotides. In particular, the present invention is directed to assays and methods of assaying oligonucleotides using one or more non-standard bases. Although the present invention is not so limited, an appreciation of various aspects of the inventions described herein will be gained through the discussion provided below. Other related assay methods for use with non-standard bases are described in U.S. Patent Provisional Application Serial No. 60/240,397, filed October 14, 2000.

[0029] As used herein, "nucleic acids" include polymeric molecules such as deoxyribonucleic acid (DNA), ribonucleic acid (RNA), peptide nucleic acid (PNA), or any sequence of what are commonly referred to as bases joined by a chemical backbone where the bases have the ability to form base pairs or hybridize with a complementary chemical structure. Suitable non-nucleotidic chemical backbones include, for example, polyamide and polymorpholino backbones. The term "nucleic acids" includes oligonucleotide, nucleotide, or polynucleotide sequences, and fragments or portions thereof. The nucleic acid can be provided in any suitable form, e.g., isolated from natural sources, recombinantly produced, or artificially synthesized, can be single- or double-stranded, and can represent the sense or antisense strand.

00977545 404504

[0030] The term "oligonucleotide" refers generally to short chain (e.g., less than about 100 nucleotides in length, and typically 6 to 50 nucleotides in length) nucleic acid sequences as prepared using techniques presently available in the art such as, for example, solid support nucleic acid synthesis, DNA replication, reverse transcription, restriction digest, run-off transcription, or the like. The exact size of the oligonucleotide will typically depend upon a variety of factors, which in turn will depend upon the ultimate function or use of the oligonucleotide.

[0031] A "sequence" refers to an ordered arrangement of nucleotides.

[0032] The term "sample" includes a specimen or culture (e.g., microbiological cultures), as well as biological samples, samples derived from biological fluids, and samples from non-biological sources.

[0033] The term "analyte" refers to a nucleic acid suspected to be in a sample. The analyte is the object of the assay (e.g., the assay determines the presence, absence, concentration, or amount of the analyte in the sample). The analyte can be directly or indirectly assayed. In at least some embodiments involving indirect assay, the analyte, if present in the sample, is used as a template to form target oligonucleotides using, for example, PCR techniques. The target oligonucleotides are then assayed to indicate the presence, absence, concentration, or amount of the analyte in the sample.

[0034] The term "target oligonucleotide" refers to oligonucleotides that are actually assayed during an assay procedure. The target oligonucleotide can be, for example, an analyte or it can be an oligonucleotide containing an analyte-specific sequence that is the same as or complementary to a sequence of the analyte. For example, the target oligonucleotide can be a product of PCR amplification of an analyte or a portion of an analyte.

[0035] The term "capture oligonucleotide" refers to an oligonucleotide having a molecular recognition sequence and coupled to a solid surface to hybridize with a target oligonucleotide having a tagging sequence or an analyte specific sequence complementary to the molecular recognition sequence, thereby capturing the target oligonucleotide on the solid surface.

[0036] A "molecular recognition sequence" as used herein is an oligonucleotide sequence complementary to the tagging sequence or to the analyte-specific sequence of a target oligonucleotide.

[0037] As used herein, the terms "complementary" or "complementarity," when used in reference to nucleic acids (i.e., a sequence of nucleotides such as an oligonucleotide or a target nucleic acid), refer to sequences that are related by base-pairing rules. For natural bases, the base pairing rules are those developed by Watson and Crick. For non-standard bases, as described herein, the base-pairing rules refer to the formation of hydrogen bonds in a manner similar to the Watson-Crick base pairing rules or the formation of specific base pairs by hydrophobic, entropic, or van der Waals forces. As an example, for the sequence "T-G-A", the complementary sequence is "A-C-T." Complementarity can be "partial," in which only some of the bases of the nucleic acids are matched according to the base pairing rules. Alternatively, there can be "complete" or "total" complementarity between the nucleic acids. The degree of complementarity between the nucleic acid strands affects the efficiency and strength of hybridization between the nucleic acid strands.

[0038] The term "hybridization" is used in reference to the pairing of complementary nucleic acids. Hybridization and the strength of hybridization (i.e., the strength of the association between the nucleic acids) is influenced by such factors as the degree of complementarity between the nucleic acids, stringency of the hybridization conditions involved, the melting temperature ( $T_m$ ) of the formed hybrid, and the G:C ratio within the nucleic acids.

[0039] Assays are performed to determine whether a sample includes an analyte having a particular nucleic acid sequence (or its complement). This nucleic acid sequence will be referred to as the "analyte-specific sequence". In at least some instances, the original sample is not directly assayed. Instead, the analyte, if present, is cloned or amplified (e.g., by PCR techniques) to provide an assay sample with a detectable amount of a target oligonucleotide that contains the analyte-specific sequence. Other techniques for amplification include, for example, nucleic acid sequence based amplification (NASBA, e.g., Guatelli, et al., Proc. Nat'l. Acad. Sci. **87**, 1874 (1990), incorporated herein by reference), strand displacement amplification (SDA, e.g., Walker, et al., Proc. Nat'l.

Acad. Sci. **89**, 392-96 (1992), incorporated herein by reference), ligase chain reaction (LCR, e.g., Kalin, et al., *Mutat. Res.*, **283**, 119-23 (1992), incorporated herein by reference), transcription mediated amplification (TMA, e.g., La Rocco, et al., *Eur. J. Clin. Microbiol. Infect. Dis.*, **13**, 726-31 (1994), incorporated herein by reference), and rolling circle amplification (RCA, e.g., Lizardi, et al., *Nat. Genet.*, **19**, 225-32 (1998), incorporated herein by reference). At least a portion of the target oligonucleotide typically corresponds to either a) the analyte, b) a portion of the analyte, c) a complement of the analyte, or d) a complement of a portion of the analyte. Detection of the target oligonucleotide by the assay indicates presence of the analyte in the original sample.

[0040] In general, an assay system for detecting one or more analyte-specific sequences includes a solid support (e.g., a chip, wafer, or a collection of solid particles). Capture oligonucleotides are disposed on the solid support in a manner which permits identification of the capture oligonucleotide (e.g., by position on a chip or wafer or by unique characteristic of particles to which particular capture oligonucleotides are attached). The capture oligonucleotides include a molecular recognition sequence. Different capture oligonucleotides with different molecular recognition sequences are used to detect different analyte-specific sequences. Using these different capture oligonucleotides, a single assay system can be designed to analyze a sample for multiple analyte-specific sequences.

[0041] Target oligonucleotides containing the analyte-specific sequences are brought into contact with the capture oligonucleotides. In addition to the analyte-specific sequence, the target oligonucleotides also each include a tagging sequence. A particular tagging sequence is associated with each analyte-specific sequence. The tagging sequence is generally complementary to one of the molecular recognition sequences. Thus, under hybridization conditions, the target oligonucleotides hybridize with the appropriate capture oligonucleotides. Alternatively, in certain methods of the present invention, the analyte-specific sequence may be complementary to one of the molecular recognition sequences.

[0042] The target oligonucleotide or its complement typically includes a reporter or a coupling agent for attachment of a reporter. Observation of the solid support to determine the presence or absence of the reporter associated with a particular capture



oligonucleotide indicates whether a particular analyte-specific sequence is present in the sample. Suitable reporters include, without limitation, biotin, fluorescents, chemilluminescents, digoxigenin, spin labels, radio labels, DNA cleavage moieties, chromaphors or fluoraphors. Examples of suitable coupling moieties include, but are not limited to, amines, thiols, hydrosines, alcohols or alkyl groups.

[0043] Examples of suitable assay systems are schematically illustrated in Figures 2A and 2B. In these assays, capture oligonucleotides 100a, 100b are coupled to a solid support 120, such as, for example, a single solid substrate 120a (e.g., a chip or wafer) or one of a number of solid particles 120b. Typically, at least one of the capture oligonucleotides (e.g., capture oligonucleotide 100a) has a molecular recognition sequence 102 that is complementary to a tagging sequence 112 of a target oligonucleotide 110 so that, under hybridization conditions, the target oligonucleotide 110 hybridizes to the capture oligonucleotide 100a.

[0044] Although assays can be prepared with all of the capture oligonucleotides having the same global molecular recognition sequence, typically, two or more different groups of capture oligonucleotides 100a, 100b are used. Each group of capture oligonucleotides has a different molecular recognition sequence. On a single solid substrate, each group of capture oligonucleotides are typically disposed on a particular region or regions of the substrate such that the region(s) is/are associated with a particular molecular recognition sequence. When a particle support is used, each group of capture oligonucleotides 100a, 100b is disposed on at least one group of particles 120b, 120c having a unique characteristic such that the capture oligonucleotide of a particular particle is determined from the characteristic of the particle to which it is attached. Such assays can be used to, for example, a) determine which allele is present in a sample by associating different capture oligonucleotides (and different regions of a substrate or different groups of particles) with each allele, b) assay for multiple related or unrelated oligonucleotides or c) both. As illustrated in Figures 2A and 2B, the target oligonucleotide preferentially hybridizes to a corresponding capture oligonucleotide permitting determination of the presence or absence of an analyte-specific sequence by observation of the presence or absence of a target oligonucleotide on a particular spatial position of the single support (Figure 2A) or attached to a particular group of particles (Figure 2B).

[0045] An additional component of the assay system is a reporter 130 that couples to the target oligonucleotide 110 (or its complement 120), as described below. The reporter 130 is the component of the assay that is subsequently detected by a detection technique (e.g., by colorimetric, fluorescence, electrophoretic, electrochemical, spectroscopic, chromatographic, densitometric, or radiographic techniques) to indicate the presence or concentration of the target oligonucleotide. The reporter will typically be determined by the detection technique (e.g., fluorophore reporters for fluorescent techniques and radio-labels for radiographic techniques.)

[0046] In some assays, one or both of the capture oligonucleotide and the target oligonucleotide include at least one non-standard base. The use of non-standard base(s) can improve the specificity of an assay that includes hybridization because non-standard bases preferentially hybridize to other complementary non-standard bases. The use of longer oligonucleotides can also increase the rate of specific hybridization. The hybridization of nucleic acids generally includes the sampling of about three to four bases for complete complementarity. These form nucleation sites. If a nucleation site is found, the hybridization proceeds down the strand. If the bases down the strand are not complementary, then the two strands release. Because the nucleation process takes time, the possibilities of finding a nucleation site when non-standard bases are used is reduced, thereby reducing the number of sampling steps needed to find a complete complement.

[0047] Alternatively, the non-standard bases are used to direct the addition of another non-standard base into a sequence (using, for example, PCR techniques). The added non-standard base can include a reporter or a coupling agent to which a reporter can be attached, thereby, permitting the highly selective incorporation of a reporter group for detection of the target oligonucleotide.

### **Oligonucleotides and Bases**

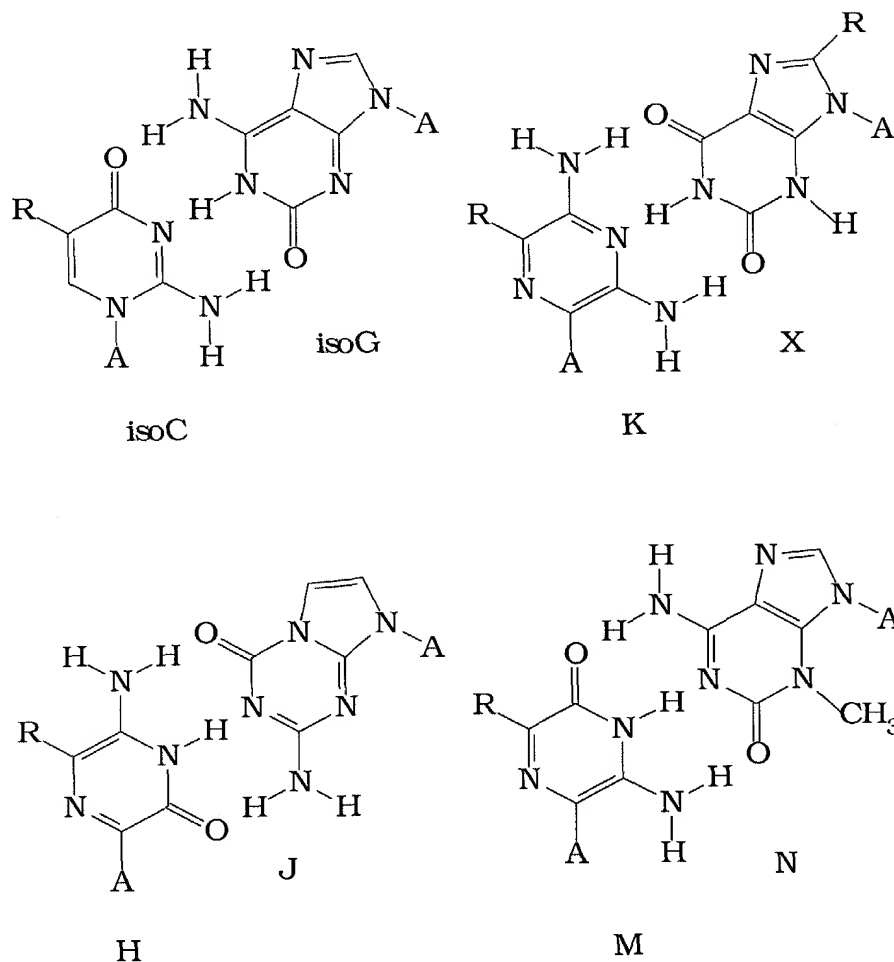
[0048] DNA and RNA are oligonucleotides that include deoxyriboses or riboses, respectively, coupled by phosphodiester bonds. Each deoxyribose or ribose includes a base coupled to a sugar. The bases incorporated in naturally-occurring DNA and RNA are adenosine (A), guanosine (G), thymidine (T), cytidine (C), and uridine (U). These five bases are "natural bases". According to the rules of base pairing elaborated by Watson and Crick, the natural bases can hybridize to form purine-pyrimidine base pairs,

where G pairs with C and A pairs with T or U. These pairing rules facilitate specific hybridization of an oligonucleotide with a complementary oligonucleotide.

[0049] The formation of these base pairs by the natural bases is facilitated by the generation of two or three hydrogen bonds between the two bases of each base pair. Each of the bases includes two or three hydrogen bond donor(s) and hydrogen bond acceptor(s). The hydrogen bonds of the base pair are each formed by the interaction of at least one hydrogen bond donor on one base with a hydrogen bond acceptor on the other base. Hydrogen bond donors include, for example, heteroatoms (e.g., oxygen or nitrogen) that have at least one attached hydrogen. Hydrogen bond acceptors include, for example, heteroatoms (e.g., oxygen or nitrogen) that have a lone pair of electrons.

[0050] The natural bases, A, G, C, T, and U, can be derivatized by substitution at non-hydrogen bonding sites to form modified natural bases. For example, a natural base can be derivatized for attachment to a support by coupling a reactive functional group (e.g., thiol, hydrazine, alcohol, or amine) to a non-hydrogen bonding atom of the base. Other possible substituents include biotin, digoxigenin, fluorescent groups, and alkyl groups (e.g., methyl or ethyl).

[0051] Non-standard bases, which form hydrogen-bonding base pairs, can also be constructed as described, for example, in U.S. Patents Nos. 5,432,272, 5,965,364, 6,001,983, and 6,037,120 and U.S. Patent Application Serial No. 08/775,401, all of which are incorporated herein by reference. By "non-standard base" it is meant a base that other than A, G, C, T, or U that is susceptible of incorporation into an oligonucleotide and which is capable of base-pairing by hydrogen bonding, or by hydrophobic, entropic, or van der Waals interactions to form base pairs with a complementary base. Figure 1 illustrates several examples of suitable bases and their corresponding base pairs. Specific examples of these bases include the following bases in base pair combinations (iso-C/iso-G, K/X, H/J, and M/N):



where A is the point of attachment to the sugar or other portion of the polymeric backbone and R is H or a substituted or unsubstituted alkyl group. It will be recognized that other non-standard bases utilizing hydrogen bonding can be prepared, as well as modifications of the above-identified non-standard bases by incorporation of functional groups at the non-hydrogen bonding atoms of the bases. To designate these non-standard bases in Figures 3 to 9, the following symbols will be used: X indicates iso-C and Y indicates iso-G.

[0052] The hydrogen bonding of these non-standard base pairs is similar to those of the natural bases where two or three hydrogen bonds are formed between hydrogen bond acceptors and hydrogen bond donors of the pairing non-standard bases. One of the differences between the natural bases and these non-standard bases is the number and position of hydrogen bond acceptors and hydrogen bond donors. For example, cytosine can be considered a donor/acceptor/acceptor base with guanine being the complementary

acceptor/donor/donor base. Iso-C is an acceptor/acceptor/donor base and iso-G is the complementary donor/donor/acceptor base, as illustrated in U.S. Patent No. 6,037,120, incorporated herein by reference.

[0053] Other non-standard bases for use in oligonucleotides include, for example, naphthalene, phenanthrene, and pyrene derivatives as discussed, for example, in Ren et al., J. Am. Chem. Soc. 118, 1671 (1996) and McMinn et al., J. Am. Chem. Soc. 121, 11585 (1999), both of which are incorporated herein by reference. These bases do not utilize hydrogen bonding for stabilization, but instead rely on hydrophobic, entropic, or van der Waals interactions to form base pairs.

### **Solid Supports**

[0054] The assay is carried out, at least in part, using a solid support. Generally, the capture oligonucleotides are coupled to or otherwise disposed on a surface of the support. A variety of different supports can be used. In some embodiments, the solid support is a single solid support, such as a chip or wafer, or the interior or exterior surface of a tube, cone, or other article. The solid support is fabricated from any suitable material to provide an optimal combination of such desired properties as stability, dimensions, shape, and surface smoothness. Preferred materials do not interfere with nucleic acid hybridization and are not subject to high amounts of non-specific binding of nucleic acids. Suitable materials include biological or nonbiological, organic or inorganic materials. For example, the master array can be fabricated from any suitable plastic or polymer, silicon, glass, ceramic, or metal, and can be provided in the form of a solid, resin, gel, rigid film, or flexible membrane. Suitable polymers include, for example, polystyrene, poly(alkyl)methacrylate, poly(vinylbenzophenone), polycarbonate, polyethylene, polypropylene, polyamide, polyvinylidene fluoride, and the like. Preferred materials include polystyrene, glass, and silicon.

[0055] In some embodiments, the single solid support 300 is divided into individual regions 310 with capture oligonucleotides disposed on the support in each region, as illustrated in Figure 3. In each of the regions or on each particle support, the capture oligonucleotides have predominantly (e.g., at least 75%) the same molecular recognition sequence. Preferably, substantially all (e.g., at least 90% and, more preferably, at least 99%) of the capture oligonucleotides have the same molecular recognition sequence in

each region or on each particle support. The capture oligonucleotides of different regions typically have different sequences, although in some instances, the same capture oligonucleotides can be used in two or more regions, for example, as a control or verification of results.

[0056] A solid support with different regions can form a regular or irregular array for testing samples and determining the presence or absence of a number of different analyte-specific sequences. For example, an array can be formed to test for 10, 100, 1000 or more different analyte-specific sequences.

[0057] Dimensions of the solid support are determined based upon such factors as the desired number of regions and the number of analyte-specific sequences to be assayed. As an example, a solid support can be provided with planar dimensions of about 0.5 cm to about 7.5 cm in length, and about 0.5 cm to about 7.5 cm in width. Solid supports can also be singly or multiply positioned on other supports, such as microscope slides (e.g., having dimensions of about 7.5 cm by about 2.5 cm). The dimensions of the solid support can be readily adapted for a particular application.

[0058] Other types of solid supports can be used. In some embodiments, the solid support is a particulate support. In these embodiments, the capture oligonucleotides are coupled to particles. Typically, the particles form groups in which particles within each group have a particular characteristic, such as, for example, color, fluorescence frequency, density, size, or shape, which can be used to distinguish or separate those particles from particles of other groups. Preferably, the particles can be separated using techniques, such as, for example, flow cytometry.

[0059] As contemplated in the invention, the particles can be fabricated from virtually any insoluble or solid material. For example, the particles can be fabricated from silica gel, glass, nylon, resins, Sephadex™, Sepharose™, cellulose, magnetic material, a metal (e.g., steel, gold, silver, aluminum, copper, or an alloy) or metal-coated material, a plastic material (e.g., polyethylene, polypropylene, polyamide, polyester, polyvinylidene fluoride (PVDF)) and the like, and combinations thereof. Examples of suitable micro-beads are described, for example, in U.S. Patents Nos. 5,736,330, 6,046,807, and 6,057,107, all of

which are incorporated herein by reference. Examples of suitable particles are available, for example, from Luminex Corp., Austin, TX.

[0060] In one embodiment, the particulate supports with associated capture oligonucleotides are disposed in a holder, such as, for example, a vial, tube, or well. The target oligonucleotide is added to the holder and the assay is conducted under hybridization conditions. The particulate supports are then separated on the basis of the unique characteristics of each group of supports. The groups of supports are then investigated to determine which support(s) have attached target oligonucleotides. Optionally, the supports can be washed to reduce the effects of cross-hybridization. One or more washes can be performed at the same or different levels of stringency, as described below. As another optional alternative, prior to contact with the support(s) and capture oligonucleotides, the solution containing target oligonucleotides can be subjected to, for example, size exclusion chromatography, differential precipitation, spin columns, or filter columns to remove primers that have not been amplified or to remove other materials that are not the same size as the target oligonucleotides.

[0061] In some embodiments, multiple holders (e.g., vials, tubes, and the like) are used to assay multiple samples or have different combinations of capture oligonucleotides (and associated supports) within each holder. As another alternative, each holder can include a single type of capture oligonucleotide (and associated support).

[0062] As another example, the support can be a group of individual support surfaces that are optionally coupled together. For example, the support can include individual optical fibers or other support members that are individually coupled to different capture oligonucleotides and then bound together to form a single article, such as a matrix.

[0063] Typically, the support (whether a single or particulate support) is capable of binding or otherwise holding the capture oligonucleotide to the surface of the support in a sufficiently stable manner to accomplish the purposes described herein. Such binding can include, for example, the formation of covalent, ionic, coordinative, hydrogen, or van der Waals bonds between the support and the capture oligonucleotides or attraction to a positively or negatively charged support. Capture oligonucleotides are attached to the solid support surface directly or via linkers. In one embodiment, capture

oligonucleotides are directly attached to the support surface by providing or derivatizing either the surface, the oligonucleotide, or both, with one or more reactive groups. For example, the surface of the Luminex<sup>TM</sup> particles can be modified with, for example, carboxylate, maleimide, or hydrazide functionalities or avidin and glass surfaces can be treated with, for example, silane or aldehyde (to form Schiff base aldehyde-amine couplings with DNA). In some embodiments, the support or a material disposed on the support (as, for example, a coating on the support) includes reactive functional groups that can couple with a reactive functional group on the capture oligonucleotides. As examples, the support can be functionalized (e.g., a metal or polymer surface that is reactively functionalized) or contain functionalities (e.g., a polymer with pending functional groups) to provide sites for coupling the capture oligonucleotides.

[0064] As an alternative, the capture oligonucleotides can be retained on the surface by cross-linking of the capture oligonucleotides. Preferably, a capture oligonucleotide that is cross-linked includes a cross-linking portion and a capture portion, where the capture portion includes a molecular recognition sequence that hybridizes to the tagging sequence of the target oligonucleotide.

[0065] As yet another alternative, the support can be partially or completely coated with a binding agent, such as streptavidin, antibody, antigen, enzyme, enzyme cofactor or inhibitor, hormone, or hormone receptor. The binding agent is typically a biological or synthetic molecule that has high affinity for another molecule or macromolecule, through covalent or non-covalent bonding. The capture oligonucleotide is coupled to a complement of the binding agent (e.g., biotin, antigen, antibody, enzyme cofactor or inhibitor, enzyme, hormone receptor, or hormone). The capture oligonucleotide is then brought in contact with the binding agent to hold the capture oligonucleotide on the support. Other known coupling techniques can be readily adapted and used in the systems and methods described herein.

### **Capture and Target Oligonucleotides**

[0066] The capture oligonucleotide includes a molecular recognition sequence that can capture, by hybridization, a target oligonucleotide having a complementary tagging sequence. The hybridization of the molecular recognition sequence of a capture oligonucleotide and the tagging sequence of a target oligonucleotide results in the



[0067] The coding and tagging sequences typically include at least six nucleotides and, in some instances, include at least 8, 10, 15, or 20 or more nucleotides. In some assays, as described below, the molecular recognition sequence and tagging sequence include one or more non-standard bases. In other assays, the molecular recognition sequence and tagging sequence do not contain non-standard bases.

[0069] In addition to the tagging sequence, the target oligonucleotide includes an analyte-specific sequence which corresponds to or is a complement to a sequence of interest in the analyte. The analyte-specific sequence can be independent from the tagging sequence or some or all of the tagging sequence can be part of the analyte-specific sequence.

[0070] The length of the capture oligonucleotides can be optimized for desired hybridization strength and kinetics. Usually, the length of the molecular recognition sequence is in the 6 to 20 (preferably, 8 to 12) nucleotide range. In a preferred embodiment, the different molecular recognition sequences of the capture oligonucleotides are not complementary to one another and, more preferably, to any known natural gene sequence or gene fragment that has a significant probability of being

present in a substantial amount in the sample to be tested. As a result, the capture molecular recognition sequences of the capture oligonucleotides will primarily hybridize to the respective complementary tagging sequences of the target oligonucleotides.

[0071] The target oligonucleotide (or an oligonucleotide complementary to at least a portion of the target oligonucleotide) includes a reporter or a coupling agent for attachment of a reporter. The reporter or coupling agent can be attached to the polymeric backbone or any of the bases of the target or complementary oligonucleotide. Techniques are known for attaching a reporter group to nucleotide bases (both natural and non-standard bases). Examples of reporter groups include biotin, digoxigenin, spin-label groups, radio labels, DNA-cleaving moieties, chromophores, and fluorophores such as fluorescein. Examples of coupling agents include biotin or substituents containing reactive functional groups. The reporter group is then provided attached to streptavidin or contains a reactive functional group that interacts with the coupling agent to bind the reporter group to the target or complementary oligonucleotide.

### **Polymerase Chain Reaction (PCR) Techniques**

[0072] A variety of Polymerase Chain Reaction (PCR) techniques are known and can be used in the assays described below. PCR techniques are typically used for the amplification of at least a portion of an oligonucleotide. The sample to be tested for the presence of an analyte-specific sequence is contacted with the first and second oligonucleotide primers; a nucleic acid polymerase; and nucleotide triphosphates corresponding to the nucleotides to be added during PCR. The natural base nucleotide triphosphates include dATP, dCTP, dGTP, dTTP, and dUTP. Nucleoside triphosphates of non-standard bases can also be added, if desired or needed. Suitable polymerases for PCR are known and include, for example, thermostable polymerases such as native and altered polymerases of *Thermus* species, including, but not limited to *Thermus aquaticus* (Taq), *Thermus flavus* (Tfl), and *Thermus thermophilus* (Tth), as well as the Klenow fragment of DNA polymerase I and the HIV-1 polymerase.

[0073] The first and second primers are complementary to different portions on different strands of the double stranded oligonucleotide that is to be amplified. The sequence of the oligonucleotide that is amplified includes the two primer sequences that hybridize to

the analyte and the region between the two primers. The primers can be formed by a variety of techniques including, for example, solid state synthesis, DNA replication, reverse transcription, restriction digest, run-off transcription, and the like.

[0074] PCR includes the cycling steps of (i) annealing the first oligonucleotide primer and the second oligonucleotide primer to the double stranded oligonucleotide that is to be amplified or to extension products formed in previous cycles; (ii) extending the annealed first and second oligonucleotide primers by the nucleic acid polymerase to synthesize primer extension products; and (iii) denaturing the products to obtain single stranded nucleic acids. Varieties of PCR have been developed by modifying the steps or varying conditions (e.g., time and temperature). Generally, any of these varieties of PCR can be used in the assays described below, although some may be more useful than others for particular assays.

[0075] One variety of PCR developed for some of the assays described below is "fast-shot PCR" in which primer extension times are reduced or eliminated. As used herein, the term "fast-shot polymerase chain reaction" or "fast-shot PCR" refers to PCR where the extension step, as well as the stops for the annealing and melting steps, are very short or eliminated. Typically, for this method, the 3' ends of the two primers are separated by no more than 10 bases on the template nucleic acid.

[0076] Enhanced specificity is achieved by using fast-shot PCR cycles where the extension step, as well as the stops for the annealing and melting steps, are very short or eliminated. In some embodiments, the PCR solution is rapidly cycled between about 90 to 100 °C and about 55 to 65°C with a maximum of about a one second hold at each temperature, thereby leaving the polymerase very little time to extend mismatched primers. In one embodiment, the reaction is cycled between about 95°C and about 58°C with about a one second hold at each temperature.

[0077] This rapid cycling is facilitated by generating a short PCR product by, in general, leaving a gap of about zero (0) to ten (10) bases on the template nucleic acid between the 3' bases of the first and second primers. Preferably, the primers are designed to have a T<sub>m</sub> of approximately 55 to 60°C. For some embodiments, a total of about 37 cycles is typically adequate to detect as little as 30 target oligonucleotides.

[0078] Allele specific PCR primers can be used to discriminate SNP (single nucleotide polymorphism) and other alleles. For SNP detection, these primers are designed to be complementary to each allele such that the polymorphic base of interest is positioned at or near (typically, within three or five bases) the 3' end of the first or second primer. High levels of allelic discrimination are achieved in part by the limited ability of Taq polymerase to extend a primer which has a nucleotide mismatch at its 3' end with that of the target DNA, *i.e.*, the corresponding allele to which the primer is not specific. Other polymerases can also be used.

[0079] Additionally, allelic discrimination can be obtained by placing the mismatch at other positions in the allele specific primer. These alternate positions for the nucleotide mismatch in the primer can be used to achieve selective amplification in two primary ways: 1) by simply lowering the T<sub>m</sub> (melting temperature) of the primer so that it is not hybridized on the template DNA during thermal cycling so that the polymerase can not extend the primers, or 2) by creating an unfavorable primer/template structure that the polymerase will not extend.

### **Examples of Assays**

#### Assays with Non-standard Bases in the Coding and Tagging Sequences

[0080] In one assay illustrated in Figure 4, two or more groups of capture oligonucleotides 202 are prepared. Each group of capture oligonucleotides 202 includes a unique molecular recognition sequence 204. The molecular recognition sequence of each group includes at least one (and, typically, two or more) non-standard bases (denoted by the use of dashed lines in the Figures). The use of non-standard bases substantially reduces the likelihood that the capture oligonucleotides will hybridize with sequences that include only natural bases. This will typically result in less non-specific hybridization when compared to a similar assay using oligonucleotides with only natural bases. The capture oligonucleotide also typically includes a reactive functional group for attachment to the solid support 206, although other attachment methods can be used, as described above.

[0081] The support for the assay can be, for example, a single solid support, such as, for example, a glass, metal, plastic, or inorganic chip. The capture oligonucleotides are disposed on the support and typically held by one of the methods described above (e.g.,

coupling via reactive groups on the capture oligonucleotide and support, use of a binding agent disposed on the support, or cross-linking of the capture oligonucleotides). Each of the groups is disposed in one or more unique regions of the solid support so that the region(s) can be associated with a particular capture oligonucleotide.

[0082] In another embodiment (not shown), the support for the assay is a particulate support (e.g., beads). It will be understood that any of the assays described herein can be performed on a single solid support, on a particulate support, or any other support. The particulate support is divided into groups of particles, each group of particles having a characteristic (e.g., color, shape, size, density, or other chemical or physical property) that distinguishes that group of particles from other groups. Each group of capture oligonucleotides is coupled to one or more groups of particles. This produces an association of a particular group of particles with a particular group of capture oligonucleotides, allowing the determination of the capture oligonucleotide by observation of the unique particle support characteristic.

[0083] Returning to Figure 4, the target oligonucleotide 208, if present in the assayed sample, contains an analyte-specific sequence 210 and a tagging sequence 212 complementary to the molecular recognition sequence 204 of one group of the capture oligonucleotides 202. The tagging sequence 212 contains at least one non-standard base; otherwise the tagging sequence would not be complementary to the molecular recognition sequence of the capture oligonucleotide. An oligonucleotide 214 complementary to a portion of the target oligonucleotide 208 includes a reporter 216 or a coupling agent (not shown) for attachment of a reporter.

[0084] The target oligonucleotide 208 and complementary oligonucleotide 214 can be formed by, for example, PCR amplification of an analyte containing the analyte-specific sequence or its complement. In PCR amplification, two different primers are used (as illustrated at B of Figure 4). A first primer 218 contains a sequence complementary to a first sequence on a first strand of the analyte 220. A second primer 222 contains a sequence that is the complementary to a second sequence on a second strand of the analyte 220 which is upstream or downstream of the first sequence. The analyte-specific sequence typically includes the sequence of the analyte stretching between, and including, the sequences (or complements) to which the primers hybridize. The first

primer 218 includes the tagging sequence 212 and the second primer 222 includes the reporter 216 (or a coupling agent for a reporter). Extension of the first and second primers and amplification proceeds using known PCR amplification techniques or the fast-shot PCR techniques described above to produce the target oligonucleotide 208 and complementary oligonucleotide 214 (as illustrated at C of Figure 4). Other known synthetic methods, such as, for example, solid state synthesis, DNA replication, reverse transcription and the like, can be used to form the target and complementary oligonucleotides.

[0085] Returning to the assay, the target oligonucleotide 208 is typically brought into contact with the support 206 (or a container holding a particulate support) with associated capture oligonucleotides 202. Conditions are controlled to promote selective hybridization of the tagging sequence of the target oligonucleotide with a complementary molecular recognition sequence of a capture oligonucleotide, if an appropriate capture oligonucleotide is present on the support (as illustrated at D of Figure 4). A reporter is also added (unless the complementary oligonucleotide 214 already contains the reporter) for coupling to the complementary oligonucleotide 214. Optionally, unincorporated primers can be removed prior to hybridization by techniques such as, for example, size exclusion chromatography, differential precipitation, spin columns, or filter columns, or after hybridization by, for example, washing.

[0086] For assays on a planar solid support, the assay can be read by determining whether the reporter group is present at each of the individual regions on the support. The presence of the reporter group indicates that the original sample contains an analyte having the analyte-specific sequence associated with the particular tagging sequence and molecular recognition sequence for that region of the support. The absence of the reporter group suggests that the sample did not contain an analyte having the particular analyte-specific sequence.

[0087] For assays on particle supports, the particles can be separated according to the unique characteristics and then it is determined which particles have a reporter coupled to the particle via the capture and target oligonucleotides. Techniques for accomplishing the separation include, for example, flow cytometry. The presence of the reporter group indicates that the sample contains the target oligonucleotide having the analyte-specific

sequence associated with a particular tagging sequence and the molecular recognition sequence of a particular capture oligonucleotide.

[0088] The assay illustrated in Figure 4 can be adapted for use in determining the presence of alleles in a sample. For example, the assay includes allele-specific primers (either the first or second primers 218, 222 or both) corresponding to two or more alleles. Each of the allele-specific primers includes a sequence that specifically hybridizes to only one allele. The tagging sequence or reporter (or coupling agent) attached to the allele-specific primer is also specific for the allele. If the allele is present in the sample, the allele-specific primer(s) associated with that allele will extend and will be detected by either hybridizing to a complementary, allele-specific capture oligonucleotide on the support or observing an allele-specific reporter group. It will be recognized that the assay can also be used to determine the presence or absence of non-allelic analyte-specific sequences in the analyte.

[0089] This method can be used to detect SNP (single nucleotide polymorphism) alleles. Either the first or second primers will be SNP-specific. Typically, two (or more) different SNP-specific primers will be used in the assay. Preferably, the SNP-specific primers will have the SNP site positioned at or near (e.g., within three or five bases) the extendable end of the primer. "Fast-shot PCR" techniques can be useful in this SNP assay because the short extension times will substantially reduce the likelihood that non-complementary primers will extend.

[0090] Hybridization of the capture oligonucleotides and target oligonucleotides is a feature of the assays described herein. This hybridization takes place in a hybridization mixture that contains salts (e.g., sodium salts or magnesium salts), a buffer (e.g., TRIS, TAPS, BICINE, or MOPs), a non-specific blocking agent (e.g., SDS, BSA, or sheared genomic DNA), and a protecting agent (e.g., EDTA or an azide), as is used in many conventional hybridization methods. Typically, the hybridization takes place at a sodium ion (or other cation) concentration of at least 0.01 to 1.0 M and a pH of 7.0 to 8.3. Generally, this hybridization and any washing steps are performed at a temperature and salt concentration that meet desired stringency conditions for maintaining hybridization. Stringency conditions are sequence dependent. Stepwise increases in stringency conditions can be used, if desired, over several washing steps.

[0091] "Low stringency conditions" are selected to be about 10 to 15°C below the thermal melting point ( $T_m$ ) for the specific sequence at the ionic strength and pH of the hybridizing solution.  $T_m$  is the temperature (for the ionic strength, pH, and nucleic acid concentration) at which about 50% of the tagging sequences hybridize to complementary molecular recognition sequences at equilibrium.

[0092] "Moderate stringency conditions" are selected to be about 5 to 10°C below the thermal melting point ( $T_m$ ) for the specific sequence at the ionic strength and pH of the hybridizing solution.

[0093] "High stringency conditions" are selected to be no more than about 5°C below the thermal melting point ( $T_m$ ) for the specific sequence at the ionic strength and pH of the hybridizing solution.

[0094] In another assay illustrated in Figure 5, two or more groups of capture oligonucleotides 252 are prepared and placed on a support 256, as illustrated at A of Figure 5. Each group of capture oligonucleotides 252 includes a unique molecular recognition sequence 254. The molecular recognition sequence of each group includes at least one (and, typically, two or more) non-standard bases. A target oligonucleotide 258 and complementary oligonucleotide 264 can be formed by, for example, PCR amplification of an analyte containing the analyte-specific sequence or its complement. In PCR amplification, two different primers are used (as illustrated at B and C of Figure 5). A first primer 268 contains a sequence complementary to a first sequence on a first strand of the analyte 270. A second primer 272 contains a sequence that is the complementary to a second sequence on a second strand of the analyte 270 which is upstream or downstream of the first sequence. The analyte-specific sequence typically includes the sequence of the analyte stretching between, and including, the sequences (or complements) to which the primers hybridize. The first primer 268 includes the tagging sequence 262 and the second primer 272 includes the reporter 266 (or a coupling agent for a reporter).

[0095] The target oligonucleotide 258 is typically brought into contact with the support 256 (or a container holding a particulate support) with associated capture oligonucleotides 252. Conditions are controlled to promote selective hybridization of the



tagging sequence of the target oligonucleotide with a complementary molecular recognition sequence of a capture oligonucleotide, if an appropriate capture oligonucleotide is present on the support (as illustrated at D of Figure 5). A reporter is also added (unless the complementary oligonucleotide 264 already contains the reporter) for coupling to the complementary oligonucleotide 264. Optionally, unincorporated primers can be removed prior to hybridization by techniques such as, for example, size exclusion chromatography, or after hybridization by, for example, washing.

[0096] An enzyme 280 is then provided to covalently couple the complementary oligonucleotide 264 to the capture oligonucleotide 252. Suitable enzymes include ligases. Optionally, the target oligonucleotide 258 is denatured from the complementary oligonucleotide 264 and the target oligonucleotide and other components of the assay are washed away leaving the complementary oligonucleotide 264 bound to the support 256, as illustrated at E of Figure 5. The reporter 266 can then be detected.

[0097] In yet another assay illustrated in Figure 6, the target oligonucleotide 314 forms a hairpin or stem-loop structure 321, 323 (or structure other than the typical double helix). In this assay, each of the first and second primers 318, 322 includes a portion of the tagging sequence 312b or a complement to a portion of the tagging sequence 312a. In addition, one of the primers 322 has a reporter 316 (or coupling agent for a reporter) attached to the portion of the tagging sequence 312b. Using, for example, PCR techniques, the first and second primers 318, 322 amplify the analyte 320 to produce a target oligonucleotide 314 and its complement 308. The tagging sequence 312b, 313a of the target oligonucleotide 314 is distributed at both ends of the target oligonucleotide.

[0098] The target oligonucleotide 314 is denatured from its complement 308 and brought into contact with the solid support 306 having capture oligonucleotides 302 with molecular recognition sequences 304. If the molecular recognition sequence 304 of one of the capture oligonucleotides is complementary to the tagging sequence 312b, 313a of the target oligonucleotide 314, the target oligonucleotide 314 will hybridize to that capture oligonucleotide. In some embodiments, the capture oligonucleotide is divided into two parts, each part complementary with one of the parts of the tagging sequence 312b, 313a. The two parts are coupled by a linker. The linker can be additional nucleotides or any other chemical linking moiety. The target sequence of the target

oligonucleotide 314 forms at least part of a stem-loop structure 321, 323 (or structure other than an double helix). Detection is then performed as discussed above in the previous example.

[0099] In an alternative assay illustrated in Figure 7, an analyte 420 is contacted by initial primers 440, 442 each having a sequence that is complementary to a sequence of the analyte 420, as illustrated at A of Figure 7. One of the initial primers 440 also includes a coupling group 444 (e.g., biotin or a substituent containing a reactive functionality) for attachment to a substrate 450. The initial primers 440, 442 are extended using, for example, PCR techniques, as illustrated at B of Figure 7. The extended initial primers 446, 448 each include the analyte-specific sequence or its complement.

[00100] The extended initial primers 446, 448 are then brought into contact with a substrate 450 that interacts with the coupling group 444 of extended initial primer 446 to attach the extended initial primer 446 to the substrate 450, as illustrated at C of Figure 7. For example, the substrate can be coated with streptavidin and the extended initial primer include biotin.

[00101] Next, first and second primers 418, 422 are brought into contact with the extended initial primers 446, 448, as illustrated at C of Figure 7. The first primer 418 has a tagging sequence 412 and the second primer 422 has a reporter 416 (or coupling agent for a reporter). Both primers also include a sequence complementary to a section of the extended initial primers 446, 448. The assay illustrated in Figure 7 also shows that other primers 422a can be added. This is not a necessary feature of the assay, but is used to illustrate one embodiment of an assay for detecting alleles. The use of allele specific primers can be used in any of the other assays illustrated herein.

[00102] In the illustrated assay, primers 422, 422a are allele-specific primers with allele-specific reporters 416, 416a. In the illustrated example, the alleles differ by a single nucleotide, although it will be understood that other allele-specific assays with more than one nucleotide difference can be performed using these techniques. Primer 422 is extended because it is complementary to a sequence on the extended initial primer 446. Primer 422a does not extend because it is not complementary to extended initial primer

446. It will be recognized that an alternative assay includes several different allele-specific primers with allele-specific tagging sequences (as opposed to allele-specific reporters). It will also be recognized that another alternative assay includes non-allelic primers for determination of the presence or absence of non-allelic analyte-specific sequences in the analyte.

[00103] The primers 418, 422 are extended to form the target oligonucleotide 408 with the tagging sequence 412 and the complementary oligonucleotide 414 with the reporter 416 (or a coupling agent for a reporter). The target oligonucleotide 408 and complementary oligonucleotide 414 are denatured from the extended initial primers 446, 448 and brought into contact with capture oligonucleotides 402 on a solid support 406 (e.g., chip, wafer, or particles). The target oligonucleotide 414 hybridizes to a capture oligonucleotide 402 having a molecular recognition sequence 404 complementary to the tagging sequence 412. The presence or absence of particular analyte-specific sequences in the analyte is determined by observation of the presence or absence of reporter associated with each unique group of capture oligonucleotides.

[00104] In another example of an assay, a first primer 468 and a second primer 472 are brought into contact with an analyte 470 and extended to form a target oligonucleotide 458 and complementary oligonucleotide 464. In the illustrated example, the first and second primers 468, 472 are both allele-specific, but specific to different alleles. In addition to the first and second primers 468, 472, other first and second primers 469, 473 are included to amplify other alleles, if present in the sample.

[00105] The first primer 468 includes a first part 462a of a tagging sequence and the second primer 472 includes a second part 462b of the tagging sequence. One of the parts 462a, 462b includes a reporter 466 (or coupling agent for a reporter). Typically, the parts 462a, 462b of the tagging sequence will be configured so that the extension of the primers 468, 472 does not proceed through the tagging sequence. For example, the parts 462a, 462b can include a non-standard base as the base linking the part of the tagging sequence to the extendable portion of the primers 468, 472. In this embodiment, the nucleotide triphosphate of the complement of the non-standard base is not included in the PCR amplification process. Alternatively, a chemical linker can be used to couple the part of the tagging sequence to the extendable portion of the primer. Examples of

[00106] A coupling oligonucleotide 452 is provided on a support 456. The coupling oligonucleotide 452 includes parts 453a, 453b that are complementary to the parts 462a, 462b of the tagging sequence. These parts 453a, 453b are coupled by a chemical or nucleotidic linker 454 that is capable of coupling 5' (or 3') ends of two nucleotidic sequences.

### Assays in which Non-standard Bases are added by PCR Techniques

[00109] To overcome the problems associated with the incorporation of multiple labeled nucleotide residues, labeled dideoxynucleic acids have been used. Because the dideoxynucleic acid lacks a 3' hydroxyl group, the oligonucleotide is terminated at the position where the labeled dideoxynucleic acid is introduced. To determine the position of the labeled nucleotide, ladders are run to sequence the oligonucleotide.

Because the oligonucleotide is terminated at the position where the dideoxyribonucleic acid is introduced, dideoxyribonucleic acids cannot generally be used in connection with amplification of the oligonucleotide strand.

[00110] Figure 9 illustrates one type of assay, according to the invention, which includes the incorporation of a non-standard base by PCR. First and second primers 518, 522 are hybridized to analyte 520 and extended. One of the primers 522 includes a non-standard base 550 which, when extended, becomes the target oligonucleotide 508. Optionally, additional bases can be provided after the non-standard base 550. The target oligonucleotide 508 with the non-standard base 550 is then brought into contact with the solid support 506a, 506b that includes capture oligonucleotides 502a, 502b. The solid support illustrated in Figure 9 is the particulate support discussed above, however, it will be recognized that a single solid support (e.g., a chip or wafer) could also be used.

[00111] The capture oligonucleotides 502a, 502b are different and are attached to different supports 506a, 506b, respectively, so that the capture oligonucleotide can be recognized by observing the unique property of the support to which it is attached. One capture oligonucleotide 502a hybridizes with the target oligonucleotide 508. The capture oligonucleotide 502a in this embodiment has a sequence that is complementary to at least a portion of the analyte-specific sequence of the target oligonucleotide 508.

[00112] After hybridization of the target oligonucleotide 508, the capture oligonucleotide 502a is extended in a PCR solution that includes dATP, dUTP, dGTP, dCTP, and the nucleotide triphosphate of a second non-standard base (e.g., diso-GTP) 552 complementary to the non-standard base 550 on the target oligonucleotide 508. The second non-standard base 552 is labeled with a reporter 516 (or coupling agent for a reporter). As the capture oligonucleotide is extended, the second non-standard base 552 with the reporter 516 is incorporated into the extended capture oligonucleotide opposite the non-standard base 550. Thus, the presence or absence of a reporter on a particular group of particulate supports indicates the presence or absence of a particular target oligonucleotide associated with the capture oligonucleotide.

[00113] Figure 10 illustrates another assay. In this assay, the first primer 618 includes a tagging sequence 612 and the second primer 622 has a non-standard base 621 (or a

[00114] The target oligonucleotide 608 is brought into contact with the solid support 606 having capture oligonucleotides 602 with molecular recognition sequences. If one of the molecular recognition sequences is complementary to the tagging sequence 612 of the target oligonucleotide 608, the target oligonucleotide 608 will hybridize to the capture oligonucleotide 602. Detection is then performed as discussed above in the previous examples.

[00116] A second extension step is then performed, in the presence of the triphosphate of the non-standard base complementary to non-standard base 721 and at least the triphosphate of the natural base complementary to natural base 723. This natural base triphosphate is labeled with a reporter 716 (or coupling group for a reporter) and is incorporated opposite natural base 723 to form the target oligonucleotide 708.

[00117] The target oligonucleotide 708 is brought into contact with the solid support 706 having capture oligonucleotides 702 with molecular recognition sequences. If one of the molecular recognition sequences is complementary to the tagging sequence 712 of the target oligonucleotide 708, the target oligonucleotide 708 will hybridize to the capture

oligonucleotide 702. Detection is then performed as discussed above in the previous examples.

[00118] In one embodiment, allele-specific second primers are used with the same first primer. The allele-specific second primers are differentiated in the portion of the second primer that anneals to the analyte. A different natural base 723 is selected for each allele. During the second extension step, where bases are added opposite the non-standard base 721 and natural base 723, the nucleotide triphosphates of two or more natural bases are added to the extension mixture. The different nucleotide triphosphates are labeled with different reporters. Thus, if the natural base 723 can be A or C, depending on the allele, the dTTP and dGTP used in the extension step are labeled with different reporters. The identity of the reporter can be used to determine the presence of a particular, associated allele. Thus, for example, four different alleles can be simultaneously tested using this method and, with appropriate choice of reporters, can be indicated using four different colors.

#### Other Assays

[00119] In one assay illustrated in Figure 16, two or more groups of capture oligonucleotides 902 are prepared. Each group of capture oligonucleotides 902 includes a unique molecular recognition sequence 904. The molecular recognition sequence of each group optionally includes at least one or more non-standard bases. The capture oligonucleotide also typically includes a reactive functional group for attachment to a solid support 906, although other attachment methods can be used, as described above.

[00120] In one embodiment, the support for the assay is a particulate support (e.g., beads). It will be understood that any of the assays described herein can be performed on a single solid support, on a particulate support, or any other support. The particulate support is divided into groups of particles, each group of particles having a characteristic (e.g., color, shape, size, density, or other chemical or physical property) that distinguishes that group of particles from other groups. Each group of capture oligonucleotides is coupled to one or more groups of particles. This produces an association of a particular group of particles with a particular group of capture oligonucleotides, allowing the determination of the capture oligonucleotide by observation of the unique particle support characteristic.

[00121] In another embodiment (not shown), the support for the assay can be, for example, a single solid support, such as, for example, a glass, metal, plastic, or inorganic chip. The capture oligonucleotides are disposed on the support and typically held by one of the methods described above (e.g., coupling via reactive groups on the capture oligonucleotide and support, use of a binding agent disposed on the support, or cross-linking of the capture oligonucleotides). Each of the groups is disposed in one or more unique regions of the solid support so that the region(s) can be associated with a particular capture oligonucleotide.

[00122] Returning to Figure 16, the target oligonucleotide 908, if present in the assayed sample, is contacted with a first primer 909 and a second primer 911. The first and second primers 909, 911 can be allele-specific or, preferably, are not complementary to allele specific portions of the target oligonucleotide (i.e., the allele specific portions of interest are positioned within the target oligonucleotide between the regions that hybridize to the two primers). The second primer 911 also includes a non-complementary attachment region 905. This non-complementary reporter attachment region 905 optionally includes one or more non-standard bases. The target oligonucleotide 908 is amplified using the first and second primers 909, 911 and PCR techniques to obtain an amplification product 907 that includes the reporter attachment region 905.

[00123] The amplification product 907 is then contacted with allele specific primers 920a, 920b that are then extended, if the particular allele is present, using reaction conditions and reaction components similar to PCR to provide an allele specific extension product 922. Each allele specific primer 920a, 920b has an allele-specific tagging sequence 912a, 912b that is complementary to different molecular recognition sequences 904 and capture oligonucleotides 902. When extending the allele specific primers 920a, 920b, a labeled nucleotide 925 (or oligonucleotide) that is complementary to one or more bases of the attachment region 905 is provided. The labeled nucleotide 925 or oligonucleotide can include a reporter or a coupling agent, such as biotin, for attachment of a reporter.

[00124] After forming the extension product 922, contact is made with the capture oligonucleotides 902 and with a reporter 930 (unless a reporter was already attached).



The capture oligonucleotide 902 and the support 906 identify which allele(s) is/are present in the sample and the reporter provides for detection of the extension product 922. For assays on particle supports, the particles can be separated according to the unique characteristics and then it is determined which particles 906 have a reporter coupled to the particle via the capture oligonucleotide 902 and extension product 922. Techniques for accomplishing the separation include, for example, flow cytometry. The presence of the reporter group indicates that the sample contains the allele associated with a particular allele-specific tagging sequence.

#### Selection of Molecular recognition sequences

[00125] When multiple molecular recognition sequences are used to form an assay system that can detect more than one analyte-specific sequence with application of a single sample, a collection of different molecular recognition sequences is typically needed. Preferably, the molecular recognition sequences are sufficiently different to permit reliable detection of analyte-specific sequences under a desired set of stringency conditions. A variety of different methods can be used to choose the collection of molecular recognition sequences. The following is a description of some methods and criteria that can be used. The methods and criteria can be used individually or in combinations.

[00126] The following are examples of criteria that can be used in creating a collection of molecular recognition sequences: the number of bases in the sequence, the number of non-standard bases in the sequence, the number of consecutive natural bases in the sequence, the number of consecutive bases (in either the forward or reverse directions) that are the same in any two sequences, specific required sequences (e.g., GC clamps at the 3' or 5' ends or both) and the estimated or actual melting temperature. One example of a method for determining  $T_m$  is described in Peyret et al., *Biochemistry*, **38**, 3468-77 (1999), incorporated herein by reference. The non-standard bases can be estimated or accounted for using, for example, values for other bases (e.g., iso-G/iso-C can be estimated using G/C) or using experimental data such as that described below.

[00127] The following are a set of steps that can be used to form the collection of molecular recognition sequences:

- 1) Create a set of all possible oligonucleotides having a length of  $n_1$  (e.g., 8, 9, or 10 nucleotides) using the natural bases and the desired non-standard bases (e.g., iso-C, iso-G, or both).
- 2) Optionally require that the oligonucleotides have a particular subsequence (e.g., GC clamps on the 3' or 5' ends or both ends).
- 3) Remove oligonucleotides without at least  $n_2$  non-standard bases (e.g., without at least two iso-C bases) or with more than  $n_3$  non-standard bases (e.g., with more than two iso-C bases) or both (e.g., accept only oligonucleotides with exactly two iso-C bases).
- 4) Optionally remove oligonucleotides with  $n_4$  (e.g., four or five) natural bases in a row.
- 5) Select one of the remaining oligonucleotides and eliminate any of the remaining oligonucleotides that have  $n_5$  bases (e.g., five or six bases) in the same order anywhere in the oligonucleotide sequence. Repeat for each non-eliminated oligonucleotide.
- 6) Optionally select one of the remaining oligonucleotides and determine its reverse complement (e.g., the reverse complement of "ACT" is "AGT"), then eliminate any of the other oligonucleotides that have  $n_6$  consecutive bases (e.g., four or five bases) that are the same as a portion of the sequence of the reverse complement. Repeat for each non-eliminated oligonucleotide.
- 7) Optionally select only the remaining oligonucleotides that have an estimated or actual melting temperature ( $T_m$ ) within a desired temperature range, above a desired temperature limit, or below a desired temperature limit. For example, oligonucleotides can be eliminated that having a melting temperature below room temperature (about 22°C).

## EXAMPLES

### Example 1

#### **Cross-hybridization Analysis of Coding and Tagging Sequences**

[00128] The equipment used in this analysis includes Luminex® 100 and Luminex® microbeads, DNA synthesizer (Northwestern Engineering, Inc.), Spectrophotometer for spot checking synthesis yields, thin layer chromatography (TLC) (SI250F TLC plate - silica gel, JTBaker) for oligonucleotide quality control, centrifuge, sonicator (Ney Dental), Vortex Genie (Vortex), and various pipettes (2, 20, 200, and 1000 $\mu$ L).

[00129] A set of more than 100 oligonucleotides (molecular recognition sequences) and their complements (tagging sequences) were designed and synthesized. The two sets of oligonucleotides contained non-standard (isoC and isoG)(EraGen Biosciences, Inc., Madison, WI) and natural (A, G, C, and T) (Perkin-Elmer/ABI) nucleotides and were 9 to 10 bases in length. The first set of the oligonucleotides was designated as molecular recognition sequences and labeled on the five prime end with an amino modifier (C6-TFA, Glen Research). The complement sets of oligonucleotides were designated the tagging sequence and labeled on the five prime end with Cy3 (Glen Research).

[00130] The following reagents were used in coupling the molecular recognition sequence to the unique Luminex beads: 0.1mM pH4.5, 2-[N-morpholino] ethanesulfonic acid (MES) (Sigma),.

[00131] 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide-HCl (EDC) (Pierce), 0.02% (v/v) Tween (Sigma), 0.1% (w/v) SDS (Sigma).

[00132] The hybridization step included a hybridization buffer Sourav 0.5 containing 10mM Tris (Sigma), 1mM EDTA (Sigma), 200mM NaCl (Aldrich), 10mM MgCl<sub>2</sub> (Aldrich), and 1% (w/v) PEG 8000 (Sigma).

[00133] Ninety-eight of the molecular recognition sequences were diluted to 1nmol/ $\mu$ L in MES. Ninety-eight unique sets of Luminex® beads were prepared for coupling. The beads were sonicated for 20 seconds and vortexed for 10 seconds before being aliquoted. From the stock beads (1.25x10<sup>7</sup> beads/mL), 5 million beads were selected and placed in a 1.5 mL microcentrifuge tube. The beads were centrifuged at 10,000 rcf for 1 minute.

The beads were then decanted, being careful not to disturb the beads. Finally, the beads were brought to 50 $\mu$ L in MES, sonicated and vortexed. To couple the molecular recognition sequence to a distinct bead, 1 nmol of each molecular recognition sequence was added to one of the unique bead sets. Next, 1.75 $\mu$ L of a fresh EDC (20mg EDC / 1mL ddH<sub>2</sub>O) was added to the mixture, sonicated and vortexed. The mixture was then allowed to incubate at room temperature in the dark for 30 minutes, vortexing every 10 minutes. After 30 minutes, another 1.75 $\mu$ L of a fresh EDC was added and incubated for 30 minutes, vortexing every 10 minutes.

[00134] After coupling, the beads were washed by adding 400 $\mu$ L Tween-20, vortexed, centrifuged (10,000 rcf/1min) and decanted. Next 400 $\mu$ L SDS was added, centrifuged, decanted and finally brought up in 100 $\mu$ L in MES and enumerated.

[00135] The complementary oligonucleotides (the tagging sequences) were quantified and qualified using TLC and polyacrylamide gel, and diluted to a final working concentration of 50 fmol/ $\mu$ L in MOPS.

[00136] After enumeration, the Luminex® bead/molecular recognition sequences were combined into a 98 bead set(1000 beads/bead region/well) for analysis. From the 98 bead set, a 50 bead set (2500 beads/bead region/well) was created. Table 1 includes the molecular recognition sequences for the 50 bead set and Table 2 includes the molecular recognition sequences for the 98 bead set.

[00137] To setup the cross hybridization experiment, 50 femtomoles of tagging sequences (1->98) were pipetted into wells in two 96 well plates (wells 1 and 2 were used for controls). Current limitations of the Luminex® 100, trimmed the dataset to 98 tagging sequences, with 2 controls for background subtraction (no tagging sequence).

[00138] The master mix of beads (98 mix), 10 $\mu$ L/well, was then added to each well along with 31 $\mu$ L of 2X Sourav 0.5 hybridization buffer and sufficient quantity of ddH<sub>2</sub>O, to give a final volume of 62 $\mu$ L/well. The reagents were mixed well and allowed to incubate at room temperature for approximately 10 minutes. The samples were immediately analyzed by flow cytometry on the Luminex®100.

[00139] The 50 bead master mix was also run with its complementary molecular recognition sequences and tagging sequences, however the tagging sequences were at 500 fmol per well.

[00140] The resulting data is reported as Median Fluorescence Intensity(MFI) per bead for both sets. Figure 12 shows the 3D surface map graphical results of the data collected in the 98 bead master mix experiment. The Y axis represents the molecular recognition sequence and the X axis represents the tagging sequence. Figure 13 shows the 3D surface map graphical results of the data collected in the 50 bead master mix experiment.

**Table 1**

**50 Bead Molecular recognition sequences (Y=iso-C and X = iso-G)**

Bead No.	Molecular recognition sequence	Seq Id No: Bead No.	Bead No.	Molecular recognition sequence	Seq Id No:
1	GAXGTXTGTC	1	26	CXTCGCXTAC	26
2	CXGTTXTTCC	2	27	GXCXAAAAXG	27
3	GGXTTGXTAG	3	28	CXXGACXATC	28
4	CTTXGXTCTC	4	29	CCATXAGXCC	29
5	CXTCAXGAAC	5	30	GGCAXTXTGG	30
6	GTAGXTAXGC	6	31	CTXAACXGGG	31
7	GGAXGXTAAC	7	32	GGAXACGXG	32
8	CXGTATXGTG	8	33	GCGXTTAXG	33
9	CATXGGTAXG	9	34	GAGXAGXTXC	34
10	GATTXTCGXC	10	35	GXCTAAXCCG	35
11	GTTXAXGACC	11	36	GCXTGTXCAC	36
12	CXGAAXGATC	12	37	GXCAGAXTCG	37
13	CAAXTACGXC	13	38	CGTXCTAGXG	38
14	CGGXATAXAC	14	39	CGXXTAGTXG	39
15	GXAAAXXAGG	15	40	CXAGGXAACC	40
16	GTCXTAGXXC	16	41	CXAGAXGAXG	41
17	GXCCTXTAXC	17	42	CGXTGXGTC	42
18	CCXACXTGAG	18	43	CAGXCGTXAG	43
19	CTXXCAXAGG	19	44	GGCTXTGXAC	44
20	GTXGAXATGC	20	45	CCAGXGXAAG	45
21	GAAAXTGXXG	21	46	GGCXAATXGC	46
22	GCTGXAXATC	22	47	GXCTGCXGG	47
23	CGCAXATXAC	23	48	GAXCTXCGGC	48
24	CTGGXTCXAG	24	49	GTXXGAXGGG	49
25	GGAAXAXXCC	25	50	GGXXATCCXG	50

Table 2

98 Bead Molecular recognition sequences (Y=iso-C and X = iso-G)

Bead No.	Molecular recognition sequence	Seq Id No:	Bead No.	Molecular recognition sequence	Seq Id No:
1	GAXGTXTGTC	1	50	CCXXATGTXG	67
2	CXGTTXTTCC	2	51	GAGXAGXTXC	34
3	GGXTTGXTAG	3	52	GXCTAAXCCG	35
4	CTTXGXTCTC	4	53	GCXTGTXCAC	36
5	CXTCAXGAAC	5	54	GXCAGAXTCG	37
6	GXCTTCXATG	51	55	CGTXCTAGXG	38
7	GTAGXTAXGC	6	56	CGXXTAGTXG	39
8	GGAXGXTAAC	7	57	CXAGGXAACC	40
9	CXGTATXGTG	8	58	GXGGTTXXTC	68
10	CATXGGTAXG	9	59	CXAGAXGAXG	41
11	GATTXTCGXG	10	60	CGXTGXGTC	42
12	GTTXAXGACC	11	61	CAGXCGTXAG	43
13	CXTCTTXXCC	52	62	GGCTXTGXAC	44
14	CXGAAXGATC	12	63	CXCCGXAATC	69
15	CAAXTACGXG	13	64	GXXACXACAC	70
16	CTCTXAXCCC	53	65	GCXCXGTXC	71
17	CTCXTGGTXC	54	66	GXCXGGAXC	72
18	CGGXATAxAC	14	67	CGAXAGCAXC	73
19	GXAAAXXAGG	15	68	CCCAXTCCXC	74
20	GTCXTAGXXC	16	69	GTXCCXXCAG	75
21	GXCCTXTAXC	17	70	CXCCTAXCGG	76
22	CCXACXTGAG	18	71	GXGTTGXCG	77
23	CTXXCAXAGG	19	72	CXAAGXAXCG	78
24	GXCAAAXCAC	55	73	GGAGXCXXTC	79
25	GTXGAXATGC	20	74	CXGXAXGTAC	80
26	GTTXGCXTTG	56	75	GXACGAXTXG	81
27	GAAAXTGXXG	21	76	GXGCTXCATG	82
28	GCTGXAXATC	22	77	GTGXAGAGXG	83
29	CXCXTXCAAC	57	78	GCCGXCXTC	84
30	CTXXACAXXC	58	79	CAAXCGXTCG	85
31	CXACTCXACC	59	80	CACAXACXGC	86
32	GACXCAXXTG	60	81	CCAGXGXAAG	45
33	CGCAXATXAC	23	82	GGCXAATXGC	46
34	CTCXCTXACG	61	83	GXCTGCXGG	47
35	CTGGXTCXAG	24	84	GXTGGXXCG	87
36	GGAAXAXXCC	25	85	GCCXCCXGT	88
37	GTGGXCTXTC	62	86	CXAXGGTCXC	89
38	CXTCGCXTAC	26	87	CCXXGXGTG	90
39	CAXXACCXAG	63	88	GGXACXCCAG	91
40	GXCXAAAAXG	27	89	GAXCTXCXGC	48
41	GTXCXAXACC	64	90	GCCTXCXGAC	92
42	CXXGACXATC	28	91	GTXCGAXGGG	49
43	CCATXAGXCC	29	92	CXTTXCGCXG	93
44	CACXXTGXTC	65	93	GGXXATCCXG	50

Bead No.	Molecular recognition sequence	Seq Id No:	Bead No.	Molecular recognition sequence	Seq Id No:
45	GGCAXTXTGG	30	94	CXCTAXGXXG	94
46	CTXAACXGGG	31	95	CXGCXAGXG	95
47	GXTCCTXGTC	66	96	CXAGCXACGG	96
48	GGAXACGXG	32	97	GACAXGCXCC	97
49	GCGXTTAXG	33	98	GGGXCGXXA	98

### Example 2

#### **Preliminary Determination of Non-standard Base Contributions to the Nearest-Neighbor Parameters for Predicting Nucleic Acid Duplex Stability**

[00141] A Beckman DU-7500 spectrometer with temperature controller and sample carriage was utilized. Six samples can simultaneously be measured with precise temperature control. In order to cover a one hundred fold range of sample concentrations, quartz cuvettes of pathlengths 0.1cm, 0.2cm, 0.5cm and 1.0cm, were obtained from Hellma, USA. DNA were synthesized on a Model 392 DNA synthesizer from Perkin-Elmer/ABI. TLC Chromatography Tank (Fisher), and TLC plates (Si250F, JTBaker). A Savant SpeedVac was used for DNA prep, as are Sep-pak C-18 purification cartridges (Waters), UV lamp, a vortex, 10cc syringes, and various pipettors (2, 20, 200, 1000 $\mu$ L)

[00142] Oligonucleotides were synthesized from natural (A, G, C, and T) nucleotides (Perkin-Elmer/ABI) and isoC, and isoG (EraGen Biosciences, Inc., Madison, WI). The synthesized self-complementary and non-self-complementary sequences are in tables 3 and 4.

Table 3: Self-Complementary Sequences (isoG = X , isoC = Y)

3A	GGA CGT CC	Control
3B	GGA YXT CC	Tandem isoC-isoG effect
3C	GXA YXT YC	IsoC-isoG in penultimate position
3D	GGA GCT CC	Control
3E	GGA XYT CC	swapped tandem isoC-isoG effect

Table 4: Non-Self-Complementary Sequences (isoG = X , isoC = Y)

4A	SEQ ID NO: 99	5' GCC AGT TTA A 3' 3' CGG TCA AAT T 5'	control
4B	SEQ ID NO:100	5' GCC AXT TTA A 3' 3' CGG TYA AAT T 5'	Single isoC-isoG in AT, TA context
4C	SEQ ID NO:101	5' GCX AGT TTA A 3' 3' CGY TCA AAT T 5'	Single isoC-isoG in mixed GC and AT context
4D	SEQ ID NO: 102	5' GYC AGT TTA A 3' 3' CXG TCA AAT T 5'	Single isoC-isoG in mixed GC and CG context
4E	SEQ ID NO: 103	5' GYY AGT TTA A 3' 3' CXX TCA AAT T 5'	Final tandem isoC-isoG substitution

[00143] The following reagents were used in the purification of the oligonucleotides and melting experiments: TLC purification was performed by eluting for 5-6hrs with *n*-propanol/ammonia/water (55:35:10 by volume)(Chou, S.-H., Flynn, P., and Reid, B. (1989) *Biochemistry* 28, 2422-2435, incorporated herein by reference). Hybridization experiments were carried out in degassed 1x SL Buffer (1.0M NaCl (Fisher), 10mM sodium cacodylate (Fisher), 0.5mM Na<sub>2</sub>EDTA(Fisher), pH 7) (SantaLucia, J., Allawi, H., and Seneviratne, P.A., (1996) *Biochemistry* 35, 3555-3562, incorporated herein by reference).

[00144] Determination of thermodynamic parameters were obtained from melting curve data using Meltwin™ v3.0 as described in Petersheim, M., and Turner, D. H. (1983) *Biochemistry* 22, 253- 263, incorporated herein by reference.

[00145] After synthesis the oligonucleotides were deprotected in ammonia at 50• C overnight, lyophilized and purified by TLC by dissolving each sample in 175μL ddH<sub>2</sub>O and eluting for 5-6 hours. The most intense, least mobile band was visualized, scraped from the plate, and eluted three times with 3 mL ddH<sub>2</sub>O. The oligonucleotides were further desalted and purified with the Sep-pak™ columns by eluting with 30% acetonitrile, 10mM ammonium bicarbonate, pH 7(SantaLucia, J., Allawi, H., and



Seneviratne, P.A., (1996) *Biochemistry* 35, 3555-3562), and finally dried in the SpeedVac™.

[00146] Self-complementary oligonucleotides were quantified and 2.0 OD<sub>260</sub> of each was collected and re-dried in the SpeedVac™. Oligonucleotides were then diluted in series to provide a one hundred fold dilution series in 1x SL Buffer. Absorbance vs. temperature profiles were measured with the Beckman DU-7500 spectrophotometer utilizing the various custom micro-cuvettes, sample carriage and temperature controller. See tables 5 and 6 for sample dilution series. The dilution series were prepared for each of the samples of Tables 3 and 4.

Table 5: Series A

Sample	volume (μL)	Add (μL)	Place into cuvette (μL)
A1	0.0	94.5	34.5
A2	57.5	40.2	34.5
A3	63.2	44.3	34.5
A4	73.0	51.2	69.0
A5	55.2	38.5	69.0

[00147] After running samples A1-A5 the dilutions for the second series were assembled. For Series B, the remaining 24.7μL from the last sample was combined with the dilutions in cuvettes A-3, A-4 and A-5 (~172.5μL total) and an additional 345μL of 1x SL Buffer.

Table 6: Series B

Sample	volume (μL)	Add (μL)	Place into cuvette (μL)
B1	542.2	0.0	172.5
B2	369.8	230.0	172.5
B3	427.2	270.0	345.0
B4	352.5	224.0	345.0
B5	231.5	132.2	345.0

[00148] The volumes placed in the cuvettes leaving approximately 4% head space in each cuvette for thermoexpansion of the samples during the melts.

[00149] For each run the samples were further degassed and then annealed by raising the temperature to 85°C for five minutes, and then cooled to 10°C over five more minutes. To limit condensation, a blanket of dry argon was utilized at low temperatures. For series A and B, measurements were taken at 260nm and 280nm, simultaneously. Samples were heated at a constant rate from 10°C to 90°C at 1.0°C/min.

[00150] The data collected from the melting experiment were then analyzed with the Meltwin™ software by curve fit analysis of  $T_m^{-1}$  vs  $\ln(C_T)$ , where  $C_T$  is the total strand concentration and  $T_m^{-1}$  is the reciprocal melting temperature (Borer, P. N., Dengler, B., Tinoco, I., Jr., and Uhlenbeck, O. C. (1974) *J. Mol. Biol.* 86, 843- 853, incorporated herein by reference).

[00151] Non-self-complementary oligonucleotides were combined in equal molar amounts to 2.0 OD<sub>260</sub> (optical density at 260 nm) and diluted in the same manner as the self-complementary oligonucleotides dilution series in Tables 5 and 6. Similar melt data was collected and analyzed with Meltwin™ for the non-self-complementary oligonucleotides.

[00152] The resulting thermodynamic parameters determined by Meltwin™ for the self-complementary and non-self-complementary oligonucleotides are summarized in Tables 7 and 8.

Table 7: Self-Complementary Sequences Thermodynamic Data(isoC = Y , isoG = X)

		$-\Delta G_{37}$ (kcal/mol)	$-\Delta H$ (kcal/mol)	$-\Delta S$ (cal/K•mol)	$T_M(^{\circ}C)$ 1.0e-4M
1A	GGA CGT CC	8.27	53.5	145.9	52.8
1B	GGA YXT CC	9.41	57.62	155.4	58.5
1C	GXA CGT YC	10.89	66.27	178.6	63.5
1D	GGA GCT CC	8.10	51.04	138.5	52.4
1E	GGA XYT CC	9.70	57.77	155.0	60.2

Table 8: Non-Self-Complementary Sequences  
Thermodynamic Data (isoC = Y, isoG = X)

			$-\Delta G_{37}$ (kcal/mol)	$-\Delta H$ (kcal/mol)	$-\Delta S$ (cal/K•mol)	$T_M(^{\circ}\text{C})$ 1.0e-4M
4A	SEQ ID NO:99	5' GCC AGT TTA A 3' 3' CGG TCA AAT T 5'	8.43	69.22	196.0	45.8
4B	SEQ ID NO:100	5' GCC AXT TTA A 3' 3' CGG TYA AAT T 5'	9.56	56.66	151.9	54.5
4C	SEQ ID NO:101	5' GCY AGT TTA A 3' 3' CGX TCA AAT T 5'	9.36	62.98	172.9	51.6
4D	SEQ ID NO:102	5' GYC AGT TTA A 3' 3' CXG TCA AAT T 5'	9.62	54.30	144.1	55.7
4E	SEQ ID NO:103	5' GYY AGT TTA A 3' 3' CXX TCA AAT T 5'	10.59	70.19	192.2	56.0

[00153] All samples have concentration dependant  $T_M$ s and monophasic melting transitions. IsoC and isoG contributions to duplex formation appear to be substantial, adding up to an additional 5°C (Sample 3B and 4C) per isoC/isoG pair to 10°C (Sample 3C and 4E) compared to natural (A, G, C, and T) Watson-Crick oligonucleotides.

[00154] Tables 7 and 8 show some the extent of the nearest-neighbor effects that are occurring when AEGIS bases are mixed with natural DNA.

### Example 3 and Comparative Example

#### Site Gated Incorporation

First primer 5' AGAACCCTTTCCTCTTCC (SEQ ID NO:104)

Target 5' AAGAACCCTTTCCTCTTCCGATGCAGGATACTTAACAATAAATATTT  
(SEQ ID NO:105)

Second Primer CTACGTCCTATGAATTGTTATTTATAAAYAGGACAGACG 5'  
(SEQ ID NO:106 )

Y = isoCTP

[00155] The sequences of the first primer, target, and second primer are shown in SEQ ID NO:104, SEQ ID NO:105, and SEQ ID NO:106, respectively.

[00156] PCR was performed using the following mixture: 0.2  $\mu\text{M}$  first primer, 0.2  $\mu\text{M}$  second primer, 50 fM target, 50  $\mu\text{M}$  each dGTP, dATP, dTTP and dCTP, 10 mM Tris pH 8, 0.1% BSA, 0.1% Triton X-100, 0.1  $\mu\text{g}/\mu\text{l}$  degraded herring sperm DNA, 40 mM KAc,

TABLE 8

Seq  
A2

2 mM MgCl<sub>2</sub>, 1U Amplitaq Stoffel (Perkin Elmer Biosciences, Foster City, CA) in a 20 µl reaction volume. The mixture was held for 2 minutes at 95°C. Then was cycled 30 times between 95°C with a 1 second hold and 58°C with a 10 second hold. Finally, the mixture was held for 2 minutes at 58°C.

[00157] Two PCR reaction mixtures were prepared. Each PCR reaction mixture was desalted using an AutoSeq™ G-50 microspin column (Amersham Pharmacia Biotech Inc., Piscataway, NJ) to remove unincorporated dNTP's, the column buffer had been exchanged for ddH<sub>2</sub>O prior to desalting the sample. The desalted samples were adjusted to these final concentrations for the following reaction components: 10 mM Tris pH 8, 0.1% BSA, 0.1% Triton X-100, 0.1 µg/µl degraded herring sperm DNA, 40 mM KAc, 2 mM MgCl<sub>2</sub>, 1U/reaction Amplitaq Stoffel (Perkin Elmer Biosciences, Foster City, CA), and 10 µM Cy3-dTTP (NEN Life Science Products, Inc., Boston, MA) in a 25 µl reaction volume. In addition, disoGTP was added in the following concentrations: 0 µM (Comparative Example) or 40 µM (Example 3). The reaction mixtures were incubated at 68°C for 15 minutes, and 5 µl of the resulting reactions were examined by electrophoresis on a 10% denaturing polyacrylamide gel. The gel was imaged for Cy3 containing extension products using a 595 Fluorimager (Molecular Dynamics, Sunnyvale, CA).

[00158] The results (data not shown) indicated that there was no additional extension of the first primer during the final PCR step when disoGTP was not present (i.e., there was little or no misincorporation of bases opposite the iso-C of the second primer).

#### **Example 4**

##### **Synthesis of Labeled deoxyisoGuanosine 5'-Triphosphates**

[00159] For the following chemical reactions, tributylammonium pyrophosphate was purchased from Sigma; biotin *N*-hydroxysuccinimide ester, was purchased from Pierce Chemical Company; all other chemicals were purchased from Aldrich Chemical Co. or Fisher Chemical Co. and were used without further purification. Solvents were dried over 4Å molecular sieves. Reactions were carried out under dry argon in oven-dry glassware. Column chromatography was performed with silica gel (230-425 mesh).

**Abbreviations:**

Ac <sub>2</sub> O	Acetic anhydride
DMF	<i>N,N</i> -Dimethylformamide
DMAP	4,4'-Dimethylaminopyridine
DMT	4,4'-Dimethoxytrityl
Et <sub>3</sub> N	Triethylamine
MeCN	Acetonitrile
MeOH	Methyl alcohol
Tol	<i>p</i> -Toluy

**1-(*p,p'*-Dimethoxytrityl)-hexamethylenediamine (2)**

[00160] Hexamethylenediamine (10 eq., 375 mmol, 43.5 g) was coevaporated two times from pyridine and dissolved in 100 ml pyridine. DMAP (0.1 eq., 3.75 mmol, 457 mg) was added and the reaction flask placed in an ice bath. DMT-chloride (1 eq., 37.5 mmol, 12.69 g), dissolved in 100 ml pyridine, was added dropwise over 2 h. It was stirred at room temperature for 4 h, MeOH (5 ml) added, the reaction mixture concentrated and the remaining residue extracted with aqueous NaHCO<sub>3</sub>/ethyl acetate. The organic layer was washed twice with aqueous NaHCO<sub>3</sub> solution, dried and the solvent evaporated. The obtained product was used in next step without further purification.

[00161] Yield: 14.895 g (35.634 mmol, 95 %) sticky oil.

**2-Chloro-6-(6-*p,p'*-dimethoxytritylamino)hexyl)-aminopurine-2'-deoxy-3',5'-ditoluytriboside (3)**

[00162] Compound 2 (1.3 equiv., 31.916 mmol, 13.34 g) was coevaporated with DMF and dissolved in 100 ml DMF. Diisopropylethylamine (3.9 equiv., 95.748 mmol, 16.65 ml) and compound 1 (1 equiv., 24.551 mmol, 13.282 g), dissolved in 100 ml DMF, were added and it was stirred at room temperature for 3 h. It was concentrated, the residue extracted with aqueous NaHCO<sub>3</sub>/ethyl acetate, the organic layer dried and the solvent evaporated. The residue was triturated with ether twice and the obtained solid product used further after drying in vacuum without further purification.

**2-Benzoyloxy-6-(6-*p,p'*-dimethoxytritylamino)hexyl)-aminopurine-2'-deoxyriboside (4)**

[00163] Compound 3 (1 equiv., 19.23 mmol, 17.74 g) was dissolved in DMF (25 ml) and added to a solution of NaH (10 eq., 192.3 mmol, 7.69 g of a 60 % dispersion in mineral

oil) in benzylalcohol (128 mL). The reaction mixture was heated (120°C, 6 h) and then stirred at room temperature (15 h) before filtrated over Celite, the filtrate evaporated, the residue extracted (ethyl acetate/water), the organic layer washed (NaHCO<sub>3</sub>-solution), dried, the solvent evaporated and the residue triturated 5 times with ether/hexane 1:10. TLC: CHCl<sub>3</sub>/10 % MeOH R<sub>F</sub> = 0.26.

[00164] Yield: 10.280 g (13.562 mmol, 70.5 % for 2 steps) foam.

**2-Benzyloxy-6-(6-p,p'-dimethoxytritylaminoethyl)-aminopurine-2'-deoxy-5'-O-p,p'-dimethoxytritylriboside (5)**

[00165] Compound 4 (14.7388 mmol, 11.172 g) was coevaporated with pyridine, dissolved in 150 ml pyridine and DMAP (0.25 equiv., 3.6847 mmol, 450 mg) added. The flask was placed in an ice bath and DMTCl (1.5 equiv., 22.108 mmol, 7.484 g) was added slowly over 2 h. It was stirred at room temperature for 22 h, then MeOH (1 ml) added, the reaction mixture concentrated and the residue extracted (chloroform/aqueous NaHCO<sub>3</sub>). The organic layer was dried, the solvent evaporated and the residue triturated with ether/hexane 1:1 to remove the excess DMT and the insoluble solid product was dried and used further without additional purification.

[00166] Yield: 14.890 g (14.047 mmol, 95 %) light brown foam.

**2-Benzyloxy-6-(6-p,p'-dimethoxytritylaminoethyl)-aminopurine-3'-O-acetyl-2'-deoxy-5'-O-p,p'-dimethoxytritylriboside (6)**

[00167] Compound 5 (14.047 mmol, 14.89 g) was coevaporated with pyridine, dissolved in 200 ml pyridine and DMAP (0.25 equiv., 3.5117 mmol, 428 mg), Et<sub>3</sub>N (5 equiv., 70.235 mmol, 9.7 ml) and Ac<sub>2</sub>O (2.5 equiv., 35.1175 mmol, 3.582 g) were added. It was stirred at room temperature for 4.5 h, then MeOH (2 ml) added, the reaction mixture concentrated and the residue extracted (ethyl acetate/aqueous NaHCO<sub>3</sub>). The organic layer was dried, the solvent evaporated and the residue purified by column chromatography using an one step gradient of ethyl acetate/hexane/Et<sub>3</sub>N 30:60:1, then 65:35:3. Yield: 5.93 g (5.385 mmol, 38 %), yellow foam.

**2-Benzyloxy-6-(6-aminoethyl)-aminopurine-3'-O-acetyl-2'-deoxyriboside (7)**

[00168] Compound 6 (2.471 mmol, 2.723 g) was dissolved in 50 ml acetonitrile/2 ml water and Ce(NH<sub>4</sub>)<sub>2</sub>(NO<sub>3</sub>)<sub>3</sub> (0.3 equiv., 0.74 mmol, 406 mg) was added. It was refluxed

for 45 min., then another 0.15 equiv.  $\text{Ce}(\text{NH}_4)_2(\text{NO}_3)_3$  (0.37 mmol, 205 mg) added and refluxing continued for 1 h. Then, it was evaporated, the residue triturated with ether to remove the DMT, the insoluble product dried and used further without additional purification.

**2-Benzoyloxy-6-(6-trifluoroacetamidoethyl)-aminopurine-3'-O-acetyl-2'-deoxyriboside (8)**

[00169] The above obtained compound **7** (max. 5.385 mmol) was dissolved in 30 ml MeOH/50 ml ethyl trifluoroacetate/5 ml  $\text{Et}_3\text{N}$  and the reaction mixture stirred at room temperature for 21.5 h. TLC (chloroform/17.5 % MeOH):  $R_F = 0.72$ ) indicated complete conversion. It was evaporated, the residue extracted (brine/ethyl acetate), the organic layer dried, the solvent evaporated and the residue purified by silica gel column chromatography using a one step gradient of chloroform/1.5 % MeOH, then 17.5 % MeOH. Yield: 2.80 g (4.714 mmol, 87 %) foam.

**2-Benzoyloxy-6-(6-trifluoroacetamidoethyl)-aminopurine-3'-O-acetyl-5'-triphosphoryl-2'-deoxyriboside (9).**

[00170] Imidazole (61 eq., 306 mg, 4.5 mmol, recrystallised) was dissolved in acetonitrile (3.6 mL) and chilled ( $0^\circ\text{C}$ ).  $\text{POCl}_3$  (19 eq., 0.128 mL) and triethylamine (61 eq., 0.633 mL) were then added and the mixture was stirred ( $0^\circ\text{C}$ , 0.5h) before adding a portion (0.309 mL) to **8** (1 eq., 0.074 mmol, 44 mg). This mixture was stirred (r.t., 0.5 h) before adding DMF (1.5 mL) containing tributylammonium pyrophosphate (2eq., 0.16 mmol, 73 mg). The reaction was then quenched (2 mL, 10%  $\text{NH}_4\text{COO}$ ) 24 h later and lyophilized. Product was purified by anion-exchange chromatography (Dionex ProPac SAX-10) using 20% MeCN and a gradient of  $(\text{NH}_4)_2\text{CO}_3$ /20% MeCN. Collected product was repetitively lyophilized to remove excess salt. Yield 0.007 mmol (10%), white solid.

**6-(6-aminoethyl)-aminopurine-5'-triphosphoryl-2'-deoxyriboside (10).**

[00171] Compound **9** (0.007 mmol) was dissolved in methanol (2.5 mL) before adding Pd/C (10%, 5 mg) and  $\text{NH}_4\text{COO}$  (0.05 mmol, 31 mg). The suspension was refluxed (1 h) before filtering off the catalyst and evaporating the solvent. The residue was then treated with 28% ammonium hydroxide (1.5 mL, 3 h, room temp.) before the reaction was dried and the product purified by anion-exchange chromatography (Dionex ProPac

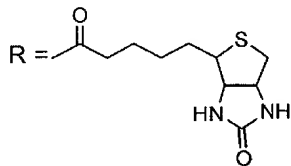
SAX-10) using 20% MeCN and a gradient of  $(\text{NH}_4)_2\text{CO}_3$ /20% MeCN. Collected product was repetitively lyophilized to remove excess salt. Yield 0.0063 mmol (90%), white solid.

**6-(6-biotinylamidohexyl)-aminopurine-5'-triphosphoryl-2'-deoxyriboside (11).**

[00172] To 10 (0.88  $\mu\text{mol}$ , triethylammonium salt) in  $\text{H}_2\text{O}$  (40  $\mu\text{L}$ ) was added sodium borate (10.5  $\mu\text{L}$ , 1M, pH 8.5) followed by DMF (216  $\mu\text{L}$ ) containing biotin *N*-hydroxysuccinimide ester (2.6  $\mu\text{mol}$ , 3 eq.). The reaction proceeded (3h, 55°C) before it was diluted with 20% MeCN and the product purified by anion-exchange chromatography (Dionex ProPac SAX-10) using  $\text{H}_2\text{O}$  and a gradient of an  $\text{NH}_4\text{HCO}_3$  solution. Yields approximately 70%.

00172





### Example 5

#### **Multiplexed genotyping of genomic DNA using incorporation of a labeled base and capture on solid support microspheres**

[00173] The genotypes of nine polymorphic loci were determined following the amplification, query, and capture of targeted nucleic acid sequences from genomic DNA samples. The first step, a multiplex PCR reaction, included a multiplexed set of paired PCR primers. Each pair of PCR primers included a first primer A and a second primer B that were designed to hybridize to and to amplify a region of mouse genomic DNA that encompasses a known polymorphic site. The second step, a multiplex allele-specific primer extension (ASPE) reaction, included a multiplexed set of tagged allele-specific primers. Each tagged allele-specific primer included a 5' tagging sequence containing non-standard nucleotides (iso-G), followed by a c3 (n-propylene) spacer, followed by a 3' sequence designed to hybridize to one of the DNA strands amplified in the previous multiplex PCR step. The allele specificity was determined by the 3' nucleotide of each tagged allele-specific primer. The multiplexed set of tagged allele-specific primers was designed to query the set of known polymorphic sites embedded in the set of multiplex PCR amplified sequences. A labeled triphosphate (dATP-Biotin) was added to the ASPE reaction, so that allele-specific extension of a tagged allele-specific primer led to the incorporation of dATP-Biotin. Unincorporated dATP-Biotin was removed prior to the subsequent capture step.

[00174] The third step, capture of the multiplex ASPE reaction products, used capture sequences containing non-standard nucleotides (iso-C) that were each covalently coupled to unique Luminex™ microsphere identities. The capture sequences were complementary to the tagging sequences used in the set of tagged allele-specific primers in the preceding ASPE reaction. Phycoerethrin was added to bind to the Biotin label on the extended tagged allele-specific primer strands and provide a fluorescent signal. Following hybridization between the capture sequences and the tagging sequences, the microspheres were injected into a Luminex100™ instrument to detect signal associated with each unique microsphere identity.

[00175] Nine polymorphic regions of the mouse genome were targeted in this example:

Target	SEQ ID NO:	Sequence	A/J	C57BL6/J
2	104	AGAAACAACCATCTAATCCCACACTAAAAT TCAAGGCTCCACAGACGAAACAGTGAAGAA TAATTGTTTCAGCATACTAACCAACTGATTA CATATTTACCATACTCAGGTTTGTGCTTCA TACAAACCCAC / TAGTCCGGCGCTCCCTGTTA GATG	CC	TT
3	105	CTTCTCCCATTGCCCAGGGCACTCTCCTCT GTAGAA / GTAGACTGATC / TTTTGTGGAGACATC A	GG	AA
4	106	AGTGCCTGCTACCTGTCAGGTGAAAATTTTC TTAGTGATCCC / TAAGCTCAATGGGTGCGYGGC TTGCAGG	CC	TT
5	107	GGTTGGAATGTTTGCACATGCAGTGTTAGT TATTTGGGC / TGATAACTACTTAGCTTATCTA GCCTGGTCCAGC	TT	CC
6	108	CTGATCTGACCTCAGACTGTTGTGCTAACA GATATAACACCAGTAAGTTGAC / GTCAAATAC TGCAGGAAGTAGAGCCTTGC	GG	CC
7	109	GACTGCTGGAGAGCTGAGGGAGGCTGTGGA GAATAAGGAGAGAGCA / GTAGTCTCGTGCCCT GCCCTGCCCATACTGAGCAGCCAAGACAC	GG	AA
8	110	GGACTGTCCAAAKGGATCTCAAGGAGAATA GTCCTTGCTATTAA / GGAGTATAAAGGCATAA AAGAGGTCATAGGGGACAACCATGACCAAG AAGTTG	AA	GG
9	111	CCTTCTGCAYTCCACAGTATAAACACAGA ATGCACACTGCA / GGTCTGTGTTTGTGTTC GATGTGAATTAAAGATGCTTTGGCTAAGCC AGGAGATGATAATACTG	AA	GG
10	112	CACATACACCATGTGAGCCATCAGCGCAAA GCCTTCGAGTTTCAGCTGTGAGATGAAGGC TTGGAGAAGCACGTTGATCTGCAAAGAAGC AAAGGAGCTAGCGGAGGCC / TGGTCACTGACC GACTGCTCA	CC	TT

The following nucleic acids were used in the multiplex PCR step for this example:

Nucleic acid component	Sequence	SEQ ID NO
PCR Primer 1A	5' -CATCTAACAGGGAGCGCC-3'	113
PCR Primer 1B	5' -6FAM-AGAAACAACCATCTAATCCCACA-3'	114
PCR Primer 2A	5' -6FAM-CTTCTCCCATTGCCCAGG-3'	115
PCR Primer 2B	5' -TGATGTCTCCACAAAGATCAGTC-3'	116
PCR Primer 3A	5' -AGTGCCTGCTACCTGTCAG-3'	117
PCR Primer 3B	5' -6FAM-CCTGCAAGCCAGCACC-3'	118
PCR Primer 4A	5' -6FAM-GGTGGAATGTTTGCACATGC-3'	119
PCR Primer 4B	5' -GCTGGACCAGGCTAGATAAGC-3'	120
PCR Primer 5A	5' -6FAM-CTGATCTGACCTCAGACTGTTG-3'	121
PCR Primer 5B	5' -GCAAGGCTCTACTTCCTGC-3'	122
PCR Primer 6A	5' -6FAM-GACTGCTGGAGAGCTGAGG-3'	123
PCR Primer 6B	5' -GTGCTTTGGCTGCTCAGTATG-3'	124
PCR Primer 7A	5' -6FAM-GGACTGTCCAAAGGGATCTC-3'	125
PCR Primer 7B	5' -CAACTTCTTGGTCATGGTTGTC-3'	126
PCR Primer 8A	5' -Cy3-CCTTCTGCAYTCCACAG-5'	127

Nucleic acid component	Sequence	SEQ ID NO
PCR Primer 8B	5' - 6FAM-CAGTATTATCATCTCCTGGCTTAGC - 3'	128
PCR Primer 9A	5' - 6FAM-CACATACACCATGTCAGCC - 3'	129
PCR Primer 9B	5' - TGAGCAGTCGGTCAGTG - 3'	130
Template 1	Mouse genomic DNA; Strain: A/J	
Template 2	Mouse genomic DNA; Strain: C57BL6/J	

[00176] PCR primers were synthesized and diluted in 1mM MOPS pH 7.5, 0.1mM EDTA. The 6FAM or Cy3 fluor on some of the PCR primers is added to allow investigation of the multiplex PCR reaction on a polyacrylamide gel.

[00177] Mouse genomic DNA samples were purchased from the Jackson Laboratory (Bar Harbor, ME). All genomic DNA samples were diluted to 5 ng/μL in 1mM MOPS pH 7.5, 0.1mM EDTA. PCR reaction components were:

Component	1X Concentration	Supplier and Location
10X PCR Buffer II	1.2X	Applied Biosystems, Foster City, CA
MgCl <sub>2</sub>	2 mM	Sigma, St. Louis, MO
dATP	200 μM	Amersham
dGTP	200 μM	Amersham
dCTP	200 μM	Amersham
dTTP	200 μM	Amersham
Amplitaq™ Gold DNA Polymerase	0.1 U/μL	Applied Biosystems, Foster City, CA
PCR Primers (each)	0.1 μM	

[00178] A Master Mix of all listed components was prepared at 1.09X concentration for 25 μL final reaction volumes. 23 μL Master Mix was combined with 2 μL of genomic DNA template (5 ng/μL) in individual PCR tubes. A negative control included water in place of genomic DNA template. PCR reactions were cycled as follows:

Cycle #	Step	Temp	Time
1	1	95°C	9 minutes
2-41	1	95°C	5 seconds
	2	55°C	30 seconds
	3	62°C	30 seconds
42	1	62°C	5 minutes
43	1	4°C	hold

[00179] Following PCR cycling, 2  $\mu$ L of each PCR reaction was transferred to act as template in the multiplex ASPE reaction. The following synthetic nucleic acids were used as tagged allele-specific (TAS) primers in the multiplex ASPE reaction:

Nucleic acid component	Sequence	SEQ ID NO
TAS Primer 1	5'-GTGYACAYGC-c3-GCTTCATACAAACCCAC-3'	131
TAS Primer 2	5'-CGAYTCTGYC-c3-GCTTCATACAAACCCAT-3'	132
TAS Primer 3	5'-CTAYCAAYCC-c3-CACTCTCCTCTGTAGAA-3'	133
TAS Primer 4	5'-GAGAYCYAAG-c3-CACTCTCCTCTGTAGAG-3'	134
TAS Primer 5	5'-GTTCYTGAYG-c3-GAAAATTTCTTAGTGATCCT-3'	135
TAS Primer 6	5'-GCTAYCTAC-c3-AAAATTTCTTAGTGATCCC-3'	136
TAS Primer 7	5'-GTTAYCYTCC-c3-AGTGTTAGTTATTTGGGT-3'	137
TAS Primer 8	5'-CACYATACYG-c3-GTGTAGTTATTTGGGC-3'	138
TAS Primer 9	5'-CYTACCYATG-c3-TAACACCAGTAAGTTGAC-3'	139
TAS Primer 10	5'-GYCGAYAATC-c3-TAACACCAGTAAGTTGAG-3'	140
TAS Primer 11	5'-GYCGTAYTTG-c3-AGAATAAGGAGAGAGCA-3'	141
TAS Primer 12	5'-GTYTATYCCG-c3-GAATAAGGAGAGAGCG-3'	142
TAS Primer 13	5'-GACAYACYTC-C3-AGAATAGTCCTTGCTATTAA-3'	143
TAS Primer 14	5'-GGAAYAACYG-C3-AGAATAGTCCTTGCTATTAG-3'	144
TAS Primer 15	5'-GATYTYCAGC-c3-AGAATGCACACTGCA-3'	145
TAS Primer 16	5'-GTYATYTGCG-c3-GAATGCACACTGCG-3'	146
TAS Primer 17	5'-GATYGTCYYG-c3-GCTAGCGGAGGCC-3'	147
TAS Primer 18	5'-GGYCTYATGG-c3-GCTAGCGGAGGCT-3'	148

[00180] Components of the ASPE reaction were:

Component	1X Concentration	Supplier and Location
Bis-Tris-Propane pH 8.9	10 mM	Sigma, St. Louis, MO
Potassium Acetate	40 mM	Sigma, St. Louis, MO
MgCl <sub>2</sub>	2 mM	Sigma, St. Louis, MO
Biotin-11-dATP,	4 $\mu$ M	NEN, Boston, MA
dGTP	200 $\mu$ M	Amersham Pharmacia, Piscataway, NJ
dCTP	200 $\mu$ M	Amersham Pharmacia, Piscataway, NJ
dTTP	200 $\mu$ M	Amersham Pharmacia, Piscataway, NJ
Amplitaq™ Gold DNA Polymerase	0.067 U/ $\mu$ L	Applied Biosystems, Foster City, CA
TAS Primers (each)	0.067 $\mu$ M	EraGen Biosciences, Inc., Madison, WI

[00181] A Master Mix containing all of the above components except TAS primers was prepared at 1.36X. Each ASPE reaction consisted of 11  $\mu$ L Master Mix, 2  $\mu$ L multiplex TAS primer mix (0.5  $\mu$ M each), 2  $\mu$ L PCR reaction (from previous step). ASPE reactions were cycled as follows:

Cycle #	Step	Temp	Time
1	1	95°C	12 minutes
2-5	1	95°C	3 seconds
	2	48°C	15 seconds
	slow ramp	30 degrees per minute	
	3	62°C	30 seconds
6	1	4°C	hold

[00182] Following ASPE cycling, 10  $\mu$ L of the reaction volume was combined with 5  $\mu$ L of a solution containing 40mM Tris, 40mM EDTA to stop the activity of the polymerase. The reaction was purified over a G-50 column to remove unincorporated dATP-Biotin. The purified multiplex ASPE reaction was then deconvoluted by capture sequences coupled to Luminex<sup>TM</sup> microspheres (Luminex Corp, Houston, TX). The coupled microspheres were:

Microsphere Identity	SEQ ID NO:	Capture Sequence
1	2	CXGTTXTTCC
2	9	CATXGGTAXG
7	14	CGGXATAxAC
15	13	CAAXTACGXC
17	22	GCTGXAXATC
18	23	CGCAXATXAC
19	1	GAXGTXTGTC
20	3	GGXTTGXTAG
21	4	CTTXGXTCTC
22	5	CXTCAXGAAC
34	7	GGAXGXTAAC
35	8	CXGTATXGTG
37	6	GTAGXTAXGC
38	10	GATTXTCGXC
45	28	CXXGACXATC
47	29	CCATXAGXCC
61	36	GCXTGTXCAC
62	37	GXCAGAXTCG

[00183] The coupled microspheres were combined in a mixture containing equal numbers of each microsphere identity in a storage buffer (10 mM MOPS pH 7.5, 200 mM NaCl, 1 mM EDTA, 1% PEG8000, 0.05% SDS). The components of the capture hybridization reaction were:

Component	1X Concentration	Supplier and Location
MOPS pH 7.5	10 mM	Fisher Chemical, Fair Lawn, NJ
NaCl	200 mM	Fisher Chemical, Fair Lawn, NJ
MgCl <sub>2</sub>	50 mM	Sigma, St. Louis, MO
EDTA	1 mM	Fisher Chemical, Fair Lawn, NJ
PEG8000	1%	Sigma, St. Louis, MO
SDS	0.05%	Fisher Chemical, Fair Lawn, NJ
Herring sperm DNA	0.1 mg/mL	Promega, Madison, WI
Microsphere mix	1000 each identity	EraGen Biosciences, Inc., Madison, WI

[00184] A Master Mix of all listed components was prepared at 1.2X concentration for 60  $\mu$ L final reaction volume. 50  $\mu$ L Master Mix was combined with 10  $\mu$ L of the purified multiplex ASPE reaction and allowed to hybridize at room temperature for 10 minutes. 10  $\mu$ L of a 0.01mg/mL solution of Streptavidin Phycoerethrin (Molecular Probes, Eugene, OR) in hybridization buffer (10mM MOPS pH7.5, 200mM NaCl, 50mM MgCl<sub>2</sub>, 1mM EDTA, 1% PEG8000, 0.05%SDS) was added to each capture hybridization reaction prior to injection into a Luminex100<sup>TM</sup> instrument.

[00185] For each capture hybridization reaction, 55  $\mu$ L was injected into the Luminex100<sup>TM</sup> at a rate of 60  $\mu$ L/min and the read continued until 50 of each microsphere identity were counted. The median fluorescence intensity was used as a measurement of the fluorescent signal associated with each microsphere identity. The results are shown in Figure 14.

### Example 6

#### **Multiplexed genotyping of genomic DNA using site-specific incorporation of a labeled non-standard base and capture on solid support microspheres**

[00186] The genotypes of nine polymorphic loci were determined following the amplification, query, and capture of targeted nucleic acid sequences from genomic DNA samples. The first step, a multiplex PCR reaction, included a multiplexed set of paired PCR primers. Each pair of PCR primers included a first primer A and a second primer B, and the second primer B included a non-standard nucleotide (iso-C) near its 5' end. Each primer pair was designed to hybridize to and to amplify a region of mouse genomic DNA that encompasses a known polymorphic site. The next step, a multiplex allele-specific primer extension (ASPE) reaction, included a multiplexed set of tagged allele-specific

primers. Each tagged allele-specific primer included a 5' tagging sequence containing non-standard nucleotides (iso-G), followed by a c3 (n-propylene) spacer, followed by a 3' sequence designed to hybridize to one of the DNA strands amplified in the previous multiplex PCR step. The allele specificity was determined by the 3' nucleotide of each tagged allele-specific primer. The multiplexed set of tagged allele-specific primers was designed to query the set of known polymorphic sites embedded in the set of multiplex PCR amplified sequences. A labeled non-standard triphosphate (isoGTP-Biotin) was added to the ASPE reaction, so that allele-specific extension of a tagged allele-specific primer led to the incorporation of the labeled non-standard triphosphate (isoGTP-Biotin) opposite the non-standard nucleotide (iso-C) in the template strand created in the preceding multiplex PCR reaction. Unincorporated isoGTP-Biotin was removed prior to the subsequent capture step.

[00187] The third step, capture of the multiplex ASPE reaction products, used capture sequences containing non-standard nucleotides (iso-C) that were each covalently coupled to unique Luminex<sup>TM</sup> microsphere identities. The capture sequences were complementary to the tagging sequences used in the set of tagged allele-specific primers in the preceding ASPE reaction. Phycoerethrin was added to bind to the Biotin label on the extended tagged allele-specific primer strands and provide fluorescent signal. Following hybridization between the capture sequences and the tagging sequences, the microspheres were injected into a Luminex100<sup>TM</sup> instrument to detect signal associated with each unique microsphere identity.

[00188] Nine polymorphic regions of the mouse genome were targeted in this example:



Target	SEQ ID NO:	Sequence	A/J	C57BL6/J	AB6F1
2	104	AGAAACAACCATCTAATCCCACACTAAAAT TCAAGGCTCCACAGACGAAACAGTGAAGAA TAATTGTTTCAGCATACTAACCAACTGATTA CATATTTACCATACTCAGGTTTGTGCTTCA TACAAACCCAC / TAGTCCGGCGCTCCCTGTTA GATG	CC	TT	CT
3	105	CTTCTCCCATTGCCCAGGGCACTCTCCTCT GTAGAA / GTAGACTGATYTTTGTGGAGACATC A	GG	AA	AG
4	106	AGTGCCTGCTACCTGTGAGGTGAAAATTTT TGTAGTATCCC / TAAGCTCAATGGGTGCGGGC TTGCAGG	CC	TT	CT
5	107	GGTTGGAATGTTTGCACATGCAGTGTAGT TATTTGGGC / TGATACTACTTAGCTTATCTA GCCTGGTCCAGC	TT	CC	CT
6	108	CTGATCTGACCTCAGACTGTTGTGCTAACA GATATAACACCAGTAAGTTGAC / GTCAAATAC TGCAGGAAGTAGAGCCTTGC	GG	CC	CG
7	109	GACTGCTGGAGAGCTGAGGGAGGCTGTGGA GAATAAGGAGAGAGCA / GTAGTCTCGTGCCC T GCCCTGCCCATACTGAGCAGCCAAGACAC	GG	AA	AG
8	110	GGACTGTCCAAKGGATCTCAAGGAGAATA GTCCTTGCTATTAA / GGAGTATAAAGGCATAA AAGAGGTCATAGGGGACAACCATGACCAAG AAGTTG	AA	GG	AG
9	111	CCTTCCTGCAYTCCACAGTATAAACACAGA ATGCACACTGCA / GGTGCTGTATTGTGTTT GATGTGAATTAAAGATGCTTTGGCTAAGCC AGGAGATGATAATACTG	AA	GG	AG
10	112	CACATACACCATGTGAGCCATCAGCGCAAA GCCTTCGAGTTTTCAGCTGTGAGATGAAGGC TTGGAGAAGCACGTTGATCTGCAAGAAGC AAAGGAGCTAGCGGAGGCC / TGGTCACTGACC GACTGCTCA	CC	TT	CT

[00189] The following nucleic acids were used in the multiplex PCR step for this example:

Nucleic acid component	Sequence	SEQ ID NO
PCR Primer 1A	5' - 6FAM-AGAAACAACCATCTAATCCCACA-3'	113
PCR Primer 1B	5' - TXCATCTAACAGGGAGCGCC-3'	114
PCR Primer 2A	5' - 6FAM-CTTCTCCCATTGCCCAGG-3'	115
PCR Primer 2B	5' - TXTGATGTCTCCACAAAGATCAGTC-3'	116
PCR Primer 3A	5' - 6FAM-CCTGCAAGCCAGCACC-3'	117
PCR Primer 3B	5' - TXCCTGCAAGCCAGCACC-3'	118
PCR Primer 4A	5' - 6FAM-GGTTGGAATGTTTGCACATGC-3'	119
PCR Primer 4B	5' - TXGCTGGACCAGGCTAGATAAGC-3'	120
PCR Primer 5A	5' - 6FAM-CTGATCTGACCTCAGACTGTTG-3'	121
PCR Primer 5B	5' - TXGCAAGGCTCTACTTCCTGC-3'	122
PCR Primer 6A	5' - 6FAM-GACTGCTGGAGAGCTGAGG-3'	123
PCR Primer 6B	5' - TXGTGTCTTGGCTGCTCAGTATG-3'	124

Nucleic acid component	Sequence	SEQ ID NO
PCR Primer 7A	5' - 6FAM-GGACTGTCCAAAGGGATCTC - 3'	125
PCR Primer 7B	5' - TXCAACTTCTTGGTCATGGTTGTC - 3'	126
PCR Primer 8A	5' - 6FAM-CAGTATTATCATCTCCTGGCTTAGC - 3'	127
PCR Primer 8B	5' - TXCCTTCCTGCACTCCACAG - 3'	128
PCR Primer 9A	5' - 6FAM-CACATACACCATGTCAGCC - 3'	129
PCR Primer 9B	5' - TXTGAGCAGTCGGTCAGTG - 3'	130
Template 1	Mouse genomic DNA; Strain: A/J	
Template 2	Mouse genomic DNA; Strain: C57BL6/J	
Template 3	Mouse genomic DNA; Strain: AB6F1	

[00190] PCR primers were synthesized and diluted in 1mM MOPS pH 7.5, 0.1mM EDTA. Mouse genomic DNA samples were purchased from the Jackson Laboratory in (Bar Harbor, ME). All genomic DNA samples were diluted to 5 ng/μL in 1mM MOPS pH 7.5, 0.1mM EDTA. PCR reaction components were:

Component	1X Concentration	Supplier and Location
10X PCR Buffer II	1.2X	Applied Biosystems, Foster City, CA
MgCl <sub>2</sub>	2 mM	Sigma, St. Louis, MO
DATP	200 μM	Amersham
DGTP	200 μM	Amersham
DCTP	200 μM	Amersham
DTTP	200 μM	Amersham
Amplitaq™ Gold DNA Polymerase	0.1 U/μL	Applied Biosystems, Foster City, CA
PCR Primers (each)	0.2 μM	

[00191] A Master Mix of all listed components was prepared at 1.09X concentration for 25 μL final reaction volumes. 23 μL Master Mix was combined with 2 μL of genomic DNA template (5 ng/μL) in individual PCR tubes. A negative control included water in place of genomic DNA template. PCR reactions were cycled as follows:

Cycle #	Step	Temp	Time
1	1	95°C	9 minutes
2-41	1	95°C	10 seconds
	2	55°C	10 seconds
	3	70°C	30 seconds
42	1	70°C	5 minutes
43	1	4°C	hold

[00192] Following PCR cycling, 2  $\mu$ L of each PCR reaction was transferred to act as template in the multiplex ASPE reaction. The following synthetic nucleic acids were used as tagged allele-specific (TAS) primers in the multiplex ASPE reaction:

Nucleic acid component	Sequence	SEQ ID NO
TAS Primer 1	5' -GTGYACAYGC-c3-GCTTCATACAAACCCAC-3'	131
TAS Primer 2	5' -CGAYTCTGYC-c3-GCTTCATACAAACCCAT-3'	132
TAS Primer 3	5' -CTAYCAAYCC-c3-CACTCTCCTCTGTAGAA-3'	133
TAS Primer 4	5' -GAGAYCYAAG-c3-CACTCTCCTCTGTAGAG-3'	134
TAS Primer 5	5' -GTTCYTGayG-c3-GAAAATTTCTTAGTGATCCT-3'	135
TAS Primer 6	5' -GCYTAYCTAC-c3-AAAATTTCTTAGTGATCCC-3'	136
TAS Primer 7	5' -GTTAYCYTCC-c3-AGTGTTAGTTATTTGGGT-3'	137
TAS Primer 8	5' -CACYATACYG-c3-GTGTAGTTATTTGGGC-3'	138
TAS Primer 9	5' -CYTACCYATG-c3-TAACACCAGTAAGTTGAC-3'	139
TAS Primer 10	5' -GYCGAYAATC-c3-TAACACCAGTAAGTTGAG-3'	140
TAS Primer 11	5' -GYCGTAYTTG-c3-AGAATAAGGAGAGAGCA-3'	141
TAS Primer 12	5' -GTYTATYCCG-c3-GAATAAGGAGAGAGCG-3'	142
TAS Primer 13	5' -GACAYACYTC-c3-AGAATAGTCCTTGCTATTAA-3'	143
TAS Primer 14	5' -GGAAYAACYG-c3-AGAATAGTCCTTGCTATTAG-3'	144
TAS Primer 15	5' -GATYTYCAGC-c3-AGAATGCACACTGCA-3'	145
TAS Primer 16	5' -GTYATYTGCG-c3-GAATGCACACTGCG-3'	146
TAS Primer 17	5' -GATYGTCTYYG-c3-GCTAGCGGAGGCC-3'	147
TAS Primer 18	5' -GGYCTYATGG-c3-GCTAGCGGAGGCT-3'	148

[00193] Components of the ASPE reaction were:

Component	1X Concentration	Supplier and Location
Bis-Tris-Propane pH 8.9	10 mM	Sigma, St. Louis, MO
Potassium Acetate	40 mM	Sigma, St. Louis, MO
MgCl <sub>2</sub>	2 mM	Sigma, St. Louis, MO
dATP	50 $\mu$ M	Amersham Pharmacia, Piscataway, NJ
dGTP	50 $\mu$ M	Amersham Pharmacia, Piscataway, NJ
dCTP	50 $\mu$ M	Amersham Pharmacia, Piscataway, NJ
dTTP	50 $\mu$ M	Amersham Pharmacia, Piscataway, NJ
d-isoGTP-Biotin	10 $\mu$ M	EraGen Biosciences, Inc., Madison, WI
Klentaq DNA Polymerase	0.067 U/ $\mu$ L	Ab Peptides, St. Louis, MO
TAS Primers (each)	0.067 $\mu$ M	EraGen Biosciences, Inc., Madison, WI

[00194] A Master Mix containing all of the above components was prepared at 1.15X. Each ASPE reaction consisted of 13  $\mu$ L Master Mix and 2  $\mu$ L PCR reaction (from previous step). ASPE reactions were cycled as follows:

Cycle #	Step	Temp	Time
1	1	95°C	2 minutes
2-11	1	95°C	1 seconds
	2	48°C	1 seconds
	3	72°C	1 minute
12	1	72°C	5 minutes
13	1	4°C	hold

[00195] Following ASPE cycling, 5  $\mu$ L of a solution containing 40mM Tris, 40mM EDTA was added to the multiplex ASPE reaction to stop the activity of the polymerase. The reaction was purified over a G-50 column to remove unincorporated d-isoGTP-Biotin. The purified multiplex ASPE reaction was then deconvoluted by capture sequences coupled to Luminex<sup>TM</sup> microspheres (Luminex Corp, Houston, TX). The coupled microspheres were:

Microsphere Identity	SEQ ID NO:	Capture Sequence
1	2	CXGTTXTTCC
2	9	CATXGGTAXG
7	14	CGGXATAXAC
15	13	CAAXTACGXC
17	22	GCTGXAXATC
18	23	CGCAXATXAC
19	1	GAXGTXTGTC
20	3	GGXTTGXTAG
21	4	CTTXGXTCTC
22	5	CXTCAXGAAC
34	7	GGAXGXTAAC
35	8	CXGTATXGTG
37	6	GTAGXTAXGC
38	10	GATTXTCGXC
45	28	CXXGACXATC
47	29	CCATXAGXCC
61	36	GCXTGTXCAC
62	37	GXCAGAXTCG

[00196] The coupled microspheres were combined in a mixture containing equal numbers of each microsphere identity in a storage buffer (10 mM MOPS pH 7.5, 200 mM NaCl, 1 mM EDTA, 1% PEG8000, 0.05% SDS). The components of the capture hybridization reaction were:

Component	1X Concentration	Supplier and Location
MOPS pH 7.5	10 mM	Fisher Chemical, Fair Lawn, NJ
NaCl	200 mM	Fisher Chemical, Fair Lawn, NJ
MgCl <sub>2</sub>	50 mM	Sigma, St. Louis, MO
EDTA	1 mM	Fisher Chemical, Fair Lawn, NJ
PEG8000	1%	Sigma, St. Louis, MO
SDS	0.05%	Fisher Chemical, Fair Lawn, NJ
Herring sperm DNA	0.1 mg/mL	Promega, Madison, WI
Microsphere mix	1000 each identity	EraGen Biosciences, Inc., Madison, WI

[00197] A Master Mix of all listed components was prepared at 1.2X concentration for 60  $\mu$ L final reaction volume. 50  $\mu$ L Master Mix was combined with 10  $\mu$ L of the purified multiplex ASPE reaction and allowed to hybridize at room temperature for 10 minutes. 10  $\mu$ L of a 0.01mg/mL solution of Streptavidin Phycoerethrin (Molecular Probes, Eugene, OR) in hybridization buffer (10mM MOPS pH7.5, 200mM NaCl, 50mM MgCl<sub>2</sub>, 1mM EDTA, 1% PEG8000, 0.05%SDS) was added to each capture hybridization reaction prior to injection into a Luminex100<sup>TM</sup> instrument.

[00198] For each capture hybridization reaction, 55  $\mu$ L was injected into the Luminex100<sup>TM</sup> at a rate of 60  $\mu$ L/min and the read continued until 50 of each microsphere identity were counted. The median fluorescence intensity was used as a measurement of the fluorescent signal associated with each microsphere identity. The results are shown in Figure 15.

[00199] The present invention should not be considered limited to the particular examples described above, but rather should be understood to cover all aspects of the invention as fairly set out in the attached claims. Various modifications, equivalent processes, as well as numerous structures to which the present invention may be applicable will be readily apparent to those of skill in the art to which the present invention is directed upon review of the instant specification.

**Example 7**

**Genotyping of genomic DNA using site specific ligation of a reporter oligonucleotide to an allele specific extension product and capture on solid support microspheres**

[00200] The genotype of a polymorphic loci was determined following the amplification, query, and capture of target nucleic acid sequences from genomic DNA samples. The first step, a PCR reaction, included a set of PCR primers: a first primer A and a second primer B. The primer B contained a 5' sequence non-complementary to the target with an iso-C at the junction of the analyte specific and non-complementary portion. The primer pair was designed to hybridize to and amplify a region of mouse genomic DNA that encompasses a known polymorphic site. The second step, a allele specific primer extension (ASPE) reaction, included a set of tagged allele-specific primers. Each tagged allele-specific primer was composed of a 5' tagging sequence containing non-standard nucleotides (iso-G), followed by a c3 spacer, followed by a 3' sequence designed to hybridize to one of the DNA strands amplified in the previous PCR step. The allele specificity was determined by the 3' nucleotide of each tagged allele-specific primer. The set of tagged allele-specific primers was designed to query a known polymorphic site embedded in the amplified sequence. A DNA ligase and a reporter oligonucleotide containing a 5' phosphate, and a 3' biotin modifications was included in the ASPE reaction. This reporter oligonucleotide was complimentary to the 5' region of primer B used to generate the amplicon that was queried. The strand of the amplified product containing this non-standard base containing region served as the template for the ASPE reaction. During allele specific primer extension, the DNA polymerase terminates at the base prior to the iso-C in the template strand, thus leaving a single stranded region to which the reporter oligonucleotide to hybridize. The complex between the extended ASPE primer, the template, and the reporter oligonucleotide results in a nick structure suitable for ligation by a DNA ligase.

[00201] The third step, capture of the multiplex ASPE reaction products, used sequences containing non-standard nucleotides (iso-C) that were each covalently coupled to unique Luminex microsphere identities. The capture sequences were complementary to the tagging sequences used in the set of tagged allele-specific primers in the preceding ASPE reaction. Streptavidin-phycoerthrin was added to bind to the biotin label on the extended and ligated allele-specific primer strands to provide fluorescent signal.

Following hybridization between the capture sequences and the tagging sequences, the microspheres were injected into a Luminex 100 instrument to detect signal associated with each unique microsphere identity.

[00202] A single polymorphic region of the mouse genome was targeted in this example:

SEQ ID NO:	Target Sequence	A/J	C57BL6/J	AB6F1
149	5'CTTCTCCCATTGCCCAGGGCACTCT CCTCTGTAGARTAGACTGATYTTTG TGGAGACATCA 3'	GG	AA	AG

Nucleic Acid Component	Sequence 5'-3'	SEQ ID NO:
PCR Primer A	PO <sub>4</sub> -CTTCTCCCATTGCCCAGG	115
PCR Primer B	CXGCXAGXGATXTGATGTCTCCACAAAGATCAGTC	150
Template 1	Mouse genomic DNA; Strain: A/J	
Template 1	Mouse genomic DNA; Strain: C57BL6/J	
Template 1	Mouse genomic DNA; Strain: AB6F1	

[00203] Mouse Genomic DNA samples were purchased from Jackson Laboratories (Bar Harbor, ME). All genomic DNA samples were diluted to 10 ng /ul in 1 mM MOPS pH 7.5, 0.1 mM EDTA. PCR reaction components were:

Component	1X Concentration	Supplier and Location
10X PCR Buffer II	1.2X	Applied Biosystems, Foster City, CA
MgCl <sub>2</sub>	2 mM	Sigma, St. Louis, MO
dGTP	200 uM	Promega, Madison, WI
dATP	200 uM	Promega, Madison, WI
dTTP	200 uM	Promega, Madison, WI
dCTP	200 uM	Promega, Madison, WI
Amplitaq DNA Polymerase Stoffel Fragment	0.2 U/ul	Applied Biosystems, Foster City, CA
PCR Primer A	0.2 uM	
PCR Primer B	0.2 uM	

[00204] A master mix of all listed components was prepared at 1.07X concentration for 30 ul final reaction volume. 23 ul of master mix was combined with 2 ul of genomic DNA template (5 ng/ul) in individual PCR tubes. A negative control included water in place of genomic DNA template. PCR reactions were thermal cycled as follows:

Cycle #	Step	Temp	Time
1	1	95°C	2 minutes
2-41	1	95°C	10 seconds
	2	55°C	10 seconds
	3	65°C	30 seconds
42	1	65°C	5 minutes
43	1	4°C	hold

[00205] Following PCR cycling, 3 ul of 5 U/ul lambda exonuclease (New England Biolabs, Beverly, MA) was added to each reaction to remove the non-template strand of the amplicon created by PCR primer A. Following addition of lambda exonuclease the reaction tubes were heated to 37°C for 5 minutes, then to 95°C for 2 minutes. Following this digest, 1 ul of each PCR reaction was transferred to act as template in the ASPE reaction. The following nucleic acid sequences were used as tagged allele-specific (TAS) primers in the ASPE reaction:

Nucleic acid component	Sequence 5'-3'	SEQ ID NO:
TAS Primer 1	CTAYCAAYCC-c3-CACTCTCCTCTGTAGAA	151
TAS Primer 2	GAGAYCYAAG-c3-CACTCTCCTCTGTAGAG	152
Reporter oligonucleotide	PO <sub>4</sub> -YATCYCTYGCYB-Biotin	153

[00206] Components of the ASPE reaction were:

Component	1X Concentration	Supplier and Location
10X PCR Buffer II	1.2X	Applied Biosystems, Foster City, CA
MgCl <sub>2</sub>	2 mM	Sigma, St. Louis, MO
DGTP	200 uM	Promega, Madison, WI
DATP	200 uM	Promega, Madison, WI
DTTP	200 uM	Promega, Madison, WI
DCTP	200 uM	Promega, Madison, WI
Amplitaq DNA Polymerase	0.1 U/ul	Applied Biosystems, Foster City, CA
Stoffel Fragment		
TAS Primer 1	0.1 uM	
TAS Primer 2	0.1 uM	
Reporter oligonucleotide	0.2 uM	
DTT	5 mM	Fisher Scientific, Pittsburgh, PA
NAD	1 mM	Roche, Indianapolis, IN
Taq DNA ligase	2 U/ul	New England Biolabs, Beverly, MA



[00207] A master mix containing all of the above components was prepared at 1.11X. Each ASPE reaction consisted of 9 ul master mix and 1 ul PCR reaction (from previous step). ASPE reactions were cycled as follows:

Cycle #	Step	Temp	Time
1	1	95°C	30 seconds
2-13	1	95°C	1 seconds
	2	48°C	1 seconds
	3	58°C	2 minutes
14	1	4°C	hold

[00208] Following ASPE cycling the reactions were deconvoluted by capture sequences coupled to Luminex microspheres (Luminex Corp, Austin, TX). The coupled microspheres were:

Microsphere Identity	SEQ ID NO:	Capture sequence 5'-3'
20	3	GGXTTGXTAG
21	4	CTTXGXTCTC

[00209] The coupled microspheres were combined in a mixture containing equal numbers of each microsphere identity in a storage buffer (10 mM MOPS pH 7.5, 200 mM NaCl, 1 mM EDTA, 1% PEG8000, 0.05% SDS), The components of the capture hybridization reaction were:

Component	1X Concentration	Supplier and Location
MOPS pH 7.5	10 mM	Sigma, St. Louis, MO
NaCl	200 mM	Sigma, St. Louis, MO
MgCl <sub>2</sub>	50 mM	Sigma, St. Louis, MO
EDTA	1 mM	Sigma, St. Louis, MO
PEG 8000	1%	Sigma, St. Louis, MO
SDS	0.05%	Sigma, St. Louis, MO
Herring Sperm DNA	0.1 mg/ml	Promega, Madison, WI
Microsphere mix	1000 each identity	Luminex Corp., Austin, TX

[00210] A master mix containing all of the above components was prepared at 1.2X concentration for a 60 ul final reaction volume. 50 ul of this master mix was added to each ASPE reaction and allowed to hybridize for 10 minutes at room temperature. 10 ul of solution of streptavidin-phycoerythrin (0.075 mg/ml in 10 mM MOPS pH 7.5, NaCl

200 mM, MgCl<sub>2</sub> 50 mM, EDTA 1 mM, PEG 8000 1%, SDS 0.05)(Molecular Probes, Eugene, OR) was added to each capture hybridization prior to injection into a Luminex 100 instrument.

[00211] For each capture hybridization reaction, 45 ul of the reaction mixture was injected into the Luminex 100 at a rate of 60 ul/min and the read continued until 100 of each microsphere identity were counted. The median fluorescence intensity (MFI) was used as a measurement of the fluorescent signal associated with each identity. The results are shown in Figure 17.

[00212] In the above example, the non-standard base of primer B at the junction of the 5' non-complementary sequence and the analyte-specific sequence is designed to prevent extension by the polymerase at the junction of the tagging and analyte specific sequences. It is specifically envisioned that other suitable linkers may be used to stop the polymerase, including, for example, 2'-O-methyl bases such as 2'-O-methyl ribonucleotides.